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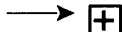


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PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

INVENTOR(S)					
Given name (first and middle [if any])		Family Name or Surname		Residence (city and either State or Foreign Country)	
John R.		Schreiber		Gates Mills, OH, USA	
<input type="checkbox"/> Additional inventors are being named on the ____ separately numbered sheets attached hereto					
TITLE OF THE INVENTION (280 characters max)					
HUMAN ANTI-PSEUDOMONAS-AERUGINOSA ANTIBODIES DERIVED FROM TRANSGENIC XENOMOUSE®					
Direct all correspondence to: CORRESPONDENCE ADDRESS					
<input checked="" type="checkbox"/> Customer Number		1473		<div>Place Customer Number Bar Code Label here</div>	
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ENCLOSED APPLICATION PARTS (check all that apply)					
<input checked="" type="checkbox"/>	Specification	Number of Pages	58	<input type="checkbox"/>	CD(s), Number
<input checked="" type="checkbox"/>	Drawing(s)	Number of Sheets	3	<input checked="" type="checkbox"/>	Other (specify)
<input checked="" type="checkbox"/>	Application Data Sheet.			<input checked="" type="checkbox"/>	Other (specify)
					Abstract
					Claims 1-109
METHOD OF PAYMENT OF FILING FEES FOR THIS PROVISIONAL APPLICATION FOR PATENT					
<input type="checkbox"/>	Applicant claims small entity status.				FILING FEE AMOUNT (\$)
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The invention was made by an agency of the United States Government or under a contract with an agency of the United States Government.					
<input type="checkbox"/>	No.				
<input checked="" type="checkbox"/>	Yes, the name of the U.S. Government agency and the Government contract number are:				
	NIH Grant Nos. AI-32596, AI-46667 and DK-27651				

Respectfully submitted,

SIGNATURE

TYPED or PRINTED NAME Grant Kalinowski

TELEPHONE 212-596-9000

Date 12/5/03

REGISTRATION NO. 48,314

Docket Number:

ABX-CW2 PROV

12/05/03

USE ONLY FOR FILING A PROVISIONAL APPLICATION FOR PATENT

This collection of information is required by 37 CFR 1.51. The information is used by the public to file (and by the PTO to process) a provisional application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 8 hours to complete, including gathering, preparing, and submitting the complete provisional application to the PTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, Washington, D.C. 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Box Provisional Application, Hon. Commissioner for Patents, Washington, D.C. 20231.

Attorney Docket No. ABX-CW2 PROV

Applicant(s) : John R. Schreiber et al.

Title : HUMAN ANTI-*PSEUDOMONAS-AERUGINOSA*
ANTIBODIES DERIVED FROM TRANSGENIC
XENOMOUSE[®]

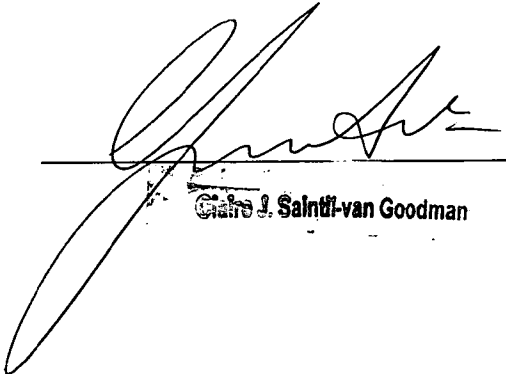
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Carlo J. Santilli-van Goodman

Encl:

- (1) Specification (58 pp.), Drawings (3 sheets)
- (2) Check for \$160.00
- (3) Print EFS Form
- (4) Provisional Application Cover Sheet
- (5) Postcard

HUMAN ANTI-*PSEUDOMONAS-AERUGINOSA* ANTIBODIES
DERIVED FROM TRANSGENIC XENOMOUSE[®]

Technical Field of the Invention

[0001] The present invention relates to compositions and methods for treating or
5 preventing *Pseudomonas aeruginosa* infection and conditions caused by such
infection. Specifically, the present invention relates to human antibodies that
specifically bind to *P. aeruginosa* lipopolysaccharide (LPS) and nucleic acid
molecules encoding them. The invention further relates to methods for making the
antibodies in a non-human animal and expressing the antibodies in cell lines including
10 hybridomas and recombinant host cell systems. The invention also relates to kits and
pharmaceutical compositions comprising the antibodies. The invention further relates
to methods of treating or preventing *P. aeruginosa* infection by administering to a
patient any of the compositions described herein.

15 **Background of the Invention**

[0002] *P. aeruginosa* are Gram-negative, flagellated rod bacteria that continue to be
a significant pathogen in nosocomial infections after surgery, prosthesis implantation
and respiratory tract procedures. *P. aeruginosa* also is an opportunistic pathogen in
the etiology of cancer, cystic fibrosis, diabetes, heart disease, otitis externa
20 (swimmer's ear), osteomyelitis, corneal ulcers, folliculitis, mastitis, pneumonia,
meningitis, urinary tract infections, endocarditis, peritonitis and other diseases found
in geriatric or immunocompromised patients.

[0003] Surgical patients are often at increased risk of *P. aeruginosa* infection by virtue of their illness (e.g., trauma, burns, inhalation injury and cancer) or treatment (e.g., disruption of natural epithelial barriers by incision or percutaneous catheterization, endotracheal intubation, cardiac and thoracic surgery, neurosurgery, and gastrointestinal surgery). Disruption of natural intestinal flora by antibiotic treatments or prophylaxis, therapeutic immunosuppression of solid organ transplant recipients, or environmental exposure to *P. aeruginosa* can place patients at increased risk. Moreover, multi-drug-resistant strains can cause significant infections in inpatient units as well as nursing homes.

10 [0004] Surgical patients are affected by nosocomial pneumonia, often caused by *P. aeruginosa*. Onset occurs after the first 72 hours of hospitalization and is characterized by fever, purulent sputum, leukocytosis and a new or changed lung infiltrate revealed by chest radiography. The oropharynx is colonized rapidly, which may spread into the lower respiratory tract. Incidence of nosocomial infection in surgical patients overall is approximately 5% to 8%, and is probably higher in all critically ill patients. The incidence of pneumonia reported from surgical intensive care units (ICUs) is 15% to 20%, and occasionally higher. See Barie et al. *Am. J. Surgery* 179:2S-7S (2000).

20 [0005] Cystic fibrosis (CF) patients suffer chronic colonization with a narrow but evolving spectrum of bacterial pathogens. *P. aeruginosa* remains the major CF pathogen with a worldwide prevalence of up to 80% to 90% in CF adults. Such infections lead to intermittent episodes of debilitating inflammatory exacerbations and progressive lung damage. Emerging pathogens also tend to be resistant to multiple antibiotic regimens. Thus, infection control plays a critical role in the quality of life and life expectancy of CF patients.

25 [0006] The onset of chronic colonization is associated with acceleration of forced expiratory volume (FEV). The original colonizing strain transforms into a mucoid colonial form which is due to copious production of a highly viscid exopolysaccharide known as alginate. The colonizing strain becomes significantly more mucinophilic and chemotactic and is associated with impaired mucociliary clearance. See Govan J. *Royal Soc. Med.* 93 Supp. 38:40-45 (2000). Moreover, the *P. aeruginosa* isolated from lungs of CF patients show changes in the LPS fatty acid acylation pattern and

enhanced resistance to the bactericidal activity of some cationic antimicrobial peptides (CAMPs).

[0007] Alterations in *P. aeruginosa* LPS lipid A were found in CF isolates that increased both bacterial resistance to antimicrobial peptides and the ability of LPS to elicit inflammatory mediators. CF patients have very high antibody titers to *P. aeruginosa* LPS in both serum and sputum, which might neutralize its biological activities *in vivo* (e.g. proinflammatory mediator release). See Pier *Trends Microbiol.* 8:247-251 (2000).

[0008] The leading cause of morbidity and mortality in severe burn wounds patients is infection with *P. aeruginosa*. See Lee et al. *Vaccine* 18:1952-1961 (2000). Burn wounds are highly exudative, creating a moist, nutrient-rich environment for bacterial colonization. Burn wounds are largely inaccessible to the patient's immune responses and vascularly delivered antibiotics due to the severe tissue injury. Moreover, burn wounds leave the host immunocompromised with endogenously decreased levels of immunoglobulin gamma (IgG). Without treatment, burn wound infections can spread and develop into sepsis with the associated production of inflammatory cytokines, including interleukin-1 (IL-1), IL-6, and tumor necrosis factors (TNFs). Burn wound infections may also result in delayed healing, increased scarring, conversion of a partial thickness defect to a full thickness defect and increased nutritional demands.

[0009] Intravenous immunoglobulin (IVIG) has been used increasingly to treat both bacterial and viral infections and primary and secondary immunodeficiency disorders. IVIG is comprised of pooled human polyclonal antibodies from normal donors which are used as a substitution therapy for primary and secondary antibody deficiencies and to treat immune-mediated diseases, including autoimmune and systemic inflammatory conditions. Immunoglobulins promote the opsonization and phagocytosis of bacteria, neutralization of bacterial toxins, inhibition of microbial attachment, and the complement-induced lysis of bacteria. See Felts et al. *Burns* 25:415-423 (1999).

[0010] Direct and local delivery of protective immunoglobulins to wound and burn sites represents a rational means to overcome the lack of vascularization of burn wounds as well as biofilm barriers. Local delivery of IgG, both prophylactically and post-infection, was demonstrated to improve survival in mouse models of *P. aeruginosa* infected burn wounds. See Felts et al. *Burns* 25:415-423 (1999).

[0011] Advances in the bioengineering of prosthetic devices has improved the lives of millions of patients. However, this progress has been tempered by implant-associated infections that often resist antibiotic treatment. Infectious organisms, including *P. aeruginosa*, preferentially target synthetic implanted materials, eliciting serious and costly infections that frequently require removal of the colonized device.

[0012] Initial microbial adhesion is a primary determinant of biomaterial colonization because initially adhering microorganisms often progress to a mature biofilm attached to the biomaterial surface. The focus of research aimed at reducing biofilm formation on prostheses has been directed toward modifying or coating the surface of the implanted materials. Approaches utilizing surface chemistry and antibiotic-releasing coatings, however, have not been fully successful.

[0013] Because surgical sites are often immunocompromised, a promising approach involves the immunostimulation of the local wound site. Studies have shown that pooled polyclonal human antibodies opsonize infecting bacteria, and pooled antibodies can inhibit *P. aeruginosa* adhesion rates and surface-growth dynamics, thus reducing biofilm formation. See Poelstra et al. *J. Biomed. Mat. Res.* **51**:224-232 (2000).

[0014] Peritonitis is often caused by ulcers, appendicitis, diverticulitis, ileus, gunshot or stab wounds, disturbances during abdominal surgery, and continuous ambulatory peritoneal dialysis (CAPD). Nosocomial peritonitis, caused by exogenous pathogenic bacteria including *P. aeruginosa*, is an especially acute problem for immunocompromised and geriatric populations.

[0015] Current treatment regimens for peritonitis focus on antibiotics. However, antibiotic resistance occurs at a significant rate and is frequently associated with clinical failure. IVIG has shown promising but inconsistent results in peritonitis, however, as with burn wounds, local (peritoneal) delivery of pooled polyclonal immunoglobulin against *P. aeruginosa* was shown to significantly reduce infection in a mouse model. See Barekzi et al. *Antimicrob. Agents Chemotherap.* **43**:1609-1615 (1999).

[0016] Treating *P. aeruginosa* infections with antibiotic regimens has become increasingly difficult because, *inter alia*, antibiotic resistant strains have arisen.

[0017] Non-human antibody preparations, including murine monoclonal antibodies, are not generally acceptable for human therapies because of their immunogenicity. Human polyclonal antibody preparations, although suitable for human therapies, have variable titers of protective antibodies for *P. aeruginosa* and a high cost of purifying antibodies from multiple donors.

[0018] Human IgM monoclonal antibodies penetrate poorly into pulmonary tissue and can be associated with immune complex formation and enhanced inflammation.

[0019] We previously described the use of immunoglobulin-inactivated mice that have been reconstituted with megabase-size contiguous fragments of human immunoglobulin loci via yeast artificial chromosomes to make entirely human monoclonal antibodies against *P. aeruginosa*. We made an IgG2 Mab against the polysaccharide (PS) portion of the LPS O-specific side chain of International Serogroup Type 06ad (Fisher Devlin It-1) *P. aeruginosa*. See International Patent Application No. WO 02/20619, published March 14, 2002, which is incorporated herein by reference in its entirety. This human Mab has strong avidity for 06ad O-side chain PS, is opsonic for uptake and killing of the bacteria by human polymorphonuclear leukocytes (PMN), and is highly protective in preventing mortality in the neutropenic mouse model of pseudomonas sepsis. However, this monoclonal antibody is highly specific for 06ad *P. aeruginosa* and therefore is not effective against other *P. aeruginosa* strains.

[0020] Therefore, there remains a need for additional therapeutically useful antibodies to treat or prevent infection with *P. aeruginosa*, methods for their preparation and use, and pharmaceutical compositions and kits comprising them.

Summary of the Invention

[0021] The present invention provides isolated antibodies, particularly human antibodies, that specifically bind to *P. aeruginosa* lipopolysaccharide (LPS) from various strains of *P. aeruginosa* and combinations of the antibodies. The invention further provides methods for making the antibodies in a non-human animal, expression of the antibodies in cell lines including hybridomas and recombinant host cell systems. The invention also provides kits and pharmaceutical compositions comprising the antibodies. Moreover, the invention provides methods of treating or

preventing pseudomonas infection by administering to a patient the pharmaceutical compositions described herein.

Brief Description of the Drawings

5 [0022] Figure 1 shows blocking of ten human Mabs to serogroup-specific *P. aeruginosa* immobilized on an ELISA plate by the corresponding purified O-side chain polysaccharide ("PS"; diamonds) but not by control PS of a different subgroup (squares).

[0023] Figure 2 depicts human complement-mediated killing of *P. aeruginosa*
10 opsonized with serogroup specific human Mabs at various doses per ml or irrelevant human Mab.

[0024] Figure 3 shows a protection experiment in neutropenic mice. The x-axis represents the serotype of the challenge strain of *P. aeruginosa* and the y-axis represents percent survival seven days after challenge.

15

Detailed Description of the Invention

[0025] In accordance with the present invention, there are provided isolated antibodies or antigen-binding portions thereof that specifically binds to *P. aeruginosa* LPS from strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4
20 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the antibodies are fully human. In some embodiments, the antibodies are monoclonal. Other preferred embodiments include nucleotide sequences encoding and amino acid sequences comprising the antibodies' heavy and light chains, and in particular sequences corresponding to a contiguous
25 heavy and light chain sequences from CDR1 through CDR3. Further provided are antibodies having similar binding properties and antibodies (or other antagonists) having similar functionality as antibodies disclosed herein. Hybridomas expressing such immunoglobulin molecules and monoclonal antibodies are also provided.

[0026] The terms herein generally have their usual meaning as understood by those
30 of ordinary skill in the art. The following terms are intended to have the following general meanings as they are used herein:

[0027] “B lymphocytic cells or progeny thereof” refer to any cell descending from, or destined for, the B lymphocytic lineage. Examples include, but are not limited to, all B lymphocytes in the B cell developmental pathway starting from the earliest B lymphocyte stem cells through memory B cells, plasma cells, and any hybridomas created *in vitro*.

[0028] “Bispecific antibodies” are single antibodies that have affinities for two separate antigens. For example, a bispecific antibody might recognize *P. aeruginosa* LPS using one combination of heavy and light chains and might recognize a leukocyte cell surface marker using a second combination of heavy and light chains attached to the first combination. See McCormick et al. *J. Immunol.* **158**:3474-3482 (1997).

[0029] “Chimeric antibodies” are antibodies that have been altered from their original form to comprise amino acid sequences from another protein. Chimeric antibodies retain at least a portion of the original antibody amino acid sequence, typically the portion comprising the antigen binding region (Fab). Examples of chimeric antibodies include, but are not limited to, bispecific antibodies and fusions with other non-immunoglobulin protein sequences.

[0030] “Cytokines” refer generally to signaling molecules of the immune system. Cytokines include, but are not limited to, Interleukins (IL), transforming growth factors (TGF), tumor necrosis factors (TNF), lymphotoxins (LT), interferons, granulocyte-macrophage colony stimulating factors (GM-CSF), macrophage CSF, Granulocyte CSF, and migration inhibition factors.

[0031] “Derivatize” refers to the process of attaching a non-immunoglobulin agent to the immunoglobulin molecules. Examples of derivatizing agents include, but are not limited to, toxins, complement, antibiotics, peptides, polysaccharides, lipids, organic polymers, radiolabels, and inorganic compounds.

[0032] “Expression control sequences” refer to sequences that allow for the inducible or constitutive expression of gene sequences under specific conditions or in specific cells. Examples of cellular processes that expression control sequence regulate include, but are not limited to, gene transcription, protein translation, messenger RNA splicing, immunoglobulin isotype switching, protein glycosylation,

protein cleavage, protein secretion, intracellular protein localization and extracellular protein homing.

[0033] “Fusion Proteins” refer to chimeric proteins comprising amino acid sequences of two or more different proteins. Typically, fusion proteins result from *in vitro* recombinatory techniques well known in the art. However, fusion proteins may result from *in vivo* crossover or other recombinatory events.

[0034] “Human immunoglobulin molecules” refer to immunoglobulin proteins that are encoded by human immunoglobulin gene sequences. The immunoglobulin gene sequences may be expressed in any non-human animal.

[0035] “Human monoclonal antibodies” refer to antibodies that are members of a population of human antibodies with identical specificities. The population of human antibodies may be produced in a hybridoma or other immortalized cell line as well as in recombinant cell lines expressing the exogenous human antibody gene sequences.

[0036] “Immunocompromised patients” refer to patients whose immune responses to foreign antigens or agents is impaired either by disease (*e.g.* AIDS), by invasive surgery, or by drug therapies in connection with treatments for other conditions (*e.g.* organ transplant patients).

[0037] “Label” refers to any molecule that attaches to the claimed immunoglobulin a functional characteristic not normally associated with that immunoglobulin. Labels can be attached via chemical modification of the immunoglobulin, recognition of the label by one of the two Fab regions of a bispecific immunoglobulin, affinity for a third agent (*e.g.* the avidin/biotin interaction), radiolabeling, or as a fusion protein expressed recombinantly. Labels can function as molecular or radioactive tags for clinical or research purposes or as agents for combating *P. aeruginosa* infection (*e.g.* toxins or complement proteins). Other examples of labels can include enzymes, fluorescent molecules, magnetic labels, epitope tags (*e.g.* *H. influenza* hemagglutinin), antibiotics, complement proteins, and cytokines.

[0038] “Respiratory patients” refer to any patient that is either being treated for a disease of the respiratory system or is receiving respiratory care, *e.g.* intubation or ventilation, in connection with some other medical treatment.

[0039] “Surgical patients” refer to any patient that is subject to an invasive surgical procedure, typically involving puncturing or incising the dermis.

[0040] “Toxins” refer to protein or non-protein compounds that can be attached to antibodies for the purpose of killing the cells to which the antibodies have attached. Examples of toxins include, but are not limited to, complement, antibiotics, peptides, polysaccharides, lipids, organic polymers, radiolabels, and inorganic compounds.

5 [0041] “Vectors” refer to DNA molecules that allow DNA sequences of interest to be cloned, propagated, recombined, mutated, or expressed outside of their native cells. Often vectors have expression control sequences that allow for the inducible or constitutive expression of gene sequences under specific conditions or in specific cells. Examples of vectors include, but are not limited to, plasmids, yeast artificial
10 chromosomes (YACs), viruses, bacteriophages, and phagemids.

[0042] “XenoMouse™” refers to mice bearing inactivated endogenous immunoglobulin loci, rendering them incapable of expressing endogenous murine immunoglobulin, but bearing substantial portions of human immunoglobulin loci. Mice of the XenoMouse™ line are capable of somatic rearrangement of the human
15 immunoglobulin genes, hypermutation of the human variable genes, and immunoglobulin isotype switching. Therefore, the mice of the XenoMouse™ line are capable of mounting effective humoral responses to antigenic challenge utilizing the human immunoglobulin gene sequences. The resulting antibodies are fully human and can be isolated from the animals themselves, from cultured cells extracted from
20 the animals, and from hybridomas created from XenoMouse™ B lymphocytic lines or progeny thereof. Moreover, the rearranged human gene sequences encoding immunoglobulins raised against specific antigenic challenges can be isolated by recombinant means well known in the art.

25 *Antibody Structure*

[0043] The basic antibody structural unit comprises a tetramer, composed of two identical pairs of polypeptide chains, each pair having one “light” (about 25 kDa) and one “heavy” chain (about 50-70 kDa). The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily
30 responsible for antigen recognition. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function. Human light chains are classified as kappa and lambda light chains. Heavy chains are classified as

mu, delta, gamma, alpha, and epsilon, and define the antibody's isotype as IgM, IgD, IgG, IgA, and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 more amino acids. *See*

5 *generally, Fundamental Immunology* Ch. 7 (Paul, W., ed., 2nd ed. Raven Press, N.Y. (1989)) (incorporated by reference in its entirety for all purposes). The variable regions of each light/heavy chain pair form the antibody binding site. Thus, an intact IgG antibody has two binding sites. Except in bifunctional or bispecific antibodies (discussed below), the two binding sites are the same.

10 **[0044]** The chains all exhibit the same general structure of relatively conserved framework regions (FR) joined by three hypervariable regions, also called complementarity determining regions or CDRs. The CDRs from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope. From N-terminal to C-terminal, both light and heavy chains comprise the

15 domains FR1, CDR1, FR2, CDR2, FR3, CDR3 and FR4. The assignment of amino acids to each domain is in accordance with the definitions of Kabat *Sequences of Proteins of Immunological Interest* (National Institutes of Health, Bethesda, Md. (1987 and 1991)), or Chothia & Lesk *J. Mol. Biol.* **196**:901-917 (1987); Chothia et al. *Nature* **342**:878-883 (1989).

20 **[0045]** A bifunctional or bispecific antibody is an artificial hybrid antibody having two different heavy/light chain pairs and two different binding sites. Bispecific antibodies can be produced by a variety of methods including fusion of hybridomas or linking of Fab' fragments. *See, e.g.,* Songsivilai & Lachmann *Clin. Exp. Immunol.* **79**:315-321 (1990), Kostelny et al. *J. Immunol.* **148**:1547-1553 (1992). In addition,

25 bispecific antibodies may be formed as "diabodies" (Holliger et al. "'Diabodies': small bivalent and bispecific antibody fragments" *Proc. Natl. Acad. Sci. USA* **90**:6444-6448 (1993)) or "Janusins" (Traunecker et al. "Bispecific single chain molecules (Janusins) target cytotoxic lymphocytes on HIV infected cells" *EMBO J.* **10**:3655-3659 (1991) and Traunecker et al. "Janusin: new molecular design for

30 bispecific reagents" *Int. J. Cancer Suppl.* **7**:51-52 (1992)). Production of bispecific antibodies can be a relatively labor intensive process compared with production of conventional antibodies and yields and degree of purity are generally lower for

bispecific antibodies. Bispecific antibodies do not exist in the form of fragments having a single binding site (e.g., Fab, Fab', and Fv).

Human Antibodies from Non-human Animals

5 [0046] Human antibodies avoid certain of the problems associated with antibodies that possess murine or rat variable and/or constant regions. The presence of such murine or rat-derived proteins can lead to the rapid clearance of the antibodies or can lead to the generation of an immune response against the antibody by a human patient. In order to avoid the utilization of murine or rat-derived antibodies, it has
10 been postulated that one can develop humanized antibodies or generate fully human antibodies through the introduction of human antibody function into a rodent so that the rodent would produce fully human antibodies.

[0047] The ability to clone and reconstruct megabase-sized human loci in YACs and to introduce them into the mouse germline provides a powerful approach to
15 elucidating the functional components of very large or crudely mapped loci as well as generating useful models of human disease. Furthermore, the utilization of such technology for substitution of mouse loci with their human equivalents could provide unique insights into the expression and regulation of human gene products during development, their communication with other systems, and their involvement in
20 disease induction and progression.

[0048] An important practical application of such a strategy is the "humanization" of the mouse humoral immune system. Introduction of human immunoglobulin (Ig) loci into mice in which the endogenous Ig genes have been inactivated offers the opportunity to study the mechanisms underlying programmed expression and
25 assembly of antibodies as well as their role in B-cell development. Furthermore, such a strategy could provide an ideal source for production of fully human monoclonal antibodies (Mabs) — an important milestone towards fulfilling the promise of antibody therapy in human disease. Fully human antibodies are expected to minimize the immunogenic and allergic responses intrinsic to mouse or mouse-derivatized
30 Mabs and thus to increase the efficacy and safety of the administered antibodies. The use of fully human antibodies can be expected to provide a substantial advantage in the treatment of chronic and recurring human diseases, such as inflammation,

autoimmunity, cancer and bacterial infections, which potentially require repeated antibody administrations.

[0049] One approach towards this goal was to engineer mouse strains deficient in mouse antibody production with large fragments of the human Ig loci in anticipation that such mice would produce a large repertoire of human antibodies in the absence of mouse antibodies. Large human Ig fragments would preserve the large variable gene diversity as well as the proper regulation of antibody production and expression. By exploiting the mouse machinery for antibody diversification and selection and the lack of immunological tolerance to human proteins, the reproduced human antibody repertoire in these mouse strains should yield high affinity antibodies against any antigen of interest, including human antigens. Using the hybridoma technology, antigen-specific human Mabs with the desired specificity could be readily produced and selected.

[0050] This general strategy was demonstrated in connection with the generation of the first XenoMouse™ strains as published in 1994. *See* Green et al. *Nature Genetics* 7:13-21 (1994). The XenoMouse™ strains were engineered with yeast artificial chromosomes (YACs) containing 245 kb and 190 kb-sized germline configuration fragments of the human heavy chain locus and kappa light chain locus, respectively, which contained core variable and constant region sequences. *Id.* The human Ig containing YACs proved to be compatible with the mouse system for both rearrangement and expression of antibodies and were capable of substituting for the inactivated mouse Ig genes. This was demonstrated by their ability to induce B-cell development, to produce an adult-like human repertoire of fully human antibodies, and to generate antigen-specific human Mabs. These results also suggested that introduction of larger portions of the human Ig loci containing greater numbers of V genes, additional regulatory elements, and human Ig constant regions might recapitulate substantially the full repertoire that is characteristic of the human humoral response to infection and immunization. The work of Green et al. was recently extended to the introduction of greater than approximately 80% of the human antibody repertoire through introduction of megabase sized, germline configuration YAC fragments of the human heavy chain loci and kappa light chain loci, respectively, to produce XenoMouse™ mice. *See* Mendez et al. *Nature Genet.*

15:146-156 (1997), Green and Jakobovits *J. Exp. Med.* **188**:483-495 (1998), and U.S. Patent Application Serial No. 08/759,620, filed December 3, 1996, the disclosures of which are hereby incorporated by reference.

[0051] Such an approach is further discussed and delineated in U.S. Patent
5 Application Serial Nos. 07/466,008, filed January 12, 1990, 07/610,515, filed
November 8, 1990, 07/919,297, filed July 24, 1992, 07/922,649, filed July 30, 1992,
filed 08/031,801, filed March 15, 1993, 08/112,848, filed August 27, 1993,
08/234,145, filed April 28, 1994, 08/376,279, filed January 20, 1995, 08/430, 938,
April 27, 1995, 08/464,584, filed June 5, 1995, 08/464,582, filed June 5, 1995,
10 08/463,191, filed June 5, 1995, 08/462,837, filed June 5, 1995, 08/486,853, filed June
5, 1995, 08/486,857, filed June 5, 1995, 08/486,859, filed June 5, 1995, 08/462,513,
filed June 5, 1995, 08/724,752, filed October 2, 1996, and 08/759,620, filed
December 3, 1996. *See also* Mendez et al. *Nature Genet.* **15**:146-156 (1997) and
Green and Jakobovits *J. Exp. Med.* **188**:483-495 (1998). *See also* U.S. Patents
15 5,916,771, 5,939,598, 5,985,615, 5,998,209, 6,075,181, 6,091,001, 6,114,598,
6,130,364, 6,162,963 and 6,150,584. *See also* WO 91/10741, WO 94/02602, WO
96/34096, WO 96/33735, WO 98/16654, WO 98/24893, WO 98/50433, WO
99/45031, WO 99/53049, WO 00/09560, WO 00/037504, European Patent No. EP 0
463 151 B1, grant published June 12, 1996, International Patent Application No. WO
20 94/02602, published February 3, 1994, International Patent Application No. WO
96/34096, published October 31, 1996, and WO 98/24893, published June 11, 1998.
The disclosures of each of the above-cited patents, applications, and references are
hereby incorporated by reference in their entirety.

[0052] Antibodies in accordance with the present invention are preferably prepared
25 through the utilization of a transgenic mouse that has a substantial portion of the
human antibody producing genome inserted but that is rendered deficient in the
production of endogenous, murine antibodies. Such mice, then, are capable of
producing human immunoglobulin molecules and antibodies and are deficient in the
production of murine immunoglobulin molecules and antibodies. Technologies
30 utilized for achieving the same are disclosed in the patents, applications, and
references disclosed herein.

[0053] Through use of such technology, fully human monoclonal antibodies, or the antigen binding portions thereof, to *P. aeruginosa* LPS were produced. Essentially, we immunized XenoMouse™ lines of mice with heat-killed *P. aeruginosa*, recovered spleen and lymph node cells (such as B-cells) from the mice that express *P.*

5 *aeruginosa* LPS antibodies, fused such recovered cells with nonsecreting myeloma cells to prepare immortal hybridoma cell lines, and screened hybridoma cell lines to identify those that produce antibodies specific to the antigen of interest.

[0054] As will be appreciated, antibodies in accordance with the present invention can be expressed in cell lines other than hybridoma cell lines. Sequences encoding particular antibodies can be used for transformation of a suitable host cell.

10 Transformation can be by any known method for introducing polynucleotides into a host cell, including, for example, packaging the polynucleotide in a virus (or into a viral vector) and transducing a host cell with the virus (or vector) or by transfection procedures known in the art, as exemplified by U.S. Patent Nos. 4,399,216,
15 4,912,040, 4,740,461, and 4,959,455 (which patents are hereby incorporated herein by reference). The transformation procedure used depends upon the host to be transformed. Methods for introduction of heterologous polynucleotides into mammalian cells are well known in the art and include dextran-mediated transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion,
20 electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the DNA into nuclei.

[0055] Mammalian cell lines available as hosts for expression are well known in the art and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to Chinese hamster ovary
25 (CHO) cells, NS/O, HeLa cells, baby hamster kidney (BHK) cells, monkey kidney cells (COS), human hepatocellular carcinoma cells (e.g., Hep G2), and a number of other cell lines. Cell lines of particular preference are selected through determining which cell lines have high expression levels and produce antibodies with constitutive *P. aeruginosa* LPS binding properties.

30 [0056] Further, expression of antibodies of the invention (or other moieties therefrom) from production cell lines can be enhanced using a number of known techniques. For example, enhanced expression can be realized by the coamplification

expression system utilizing dihydrofolate reductase (DHFR) or the glutamine synthetase gene expression system (the GS system). *See, e.g.* Kaufman and Sharp *J. Mol. Biol.* **159**:601-621 (1982); European Patent Nos. 0 216 846, 0 256 055, and 0 323 997; and European Patent Application No. 89303964.4.

5 **[0057]** Antibodies of the invention can also be produced through the generation of a mammal or plant that is transgenic for the immunoglobulin heavy and light chain sequences of interest and production of the antibody in a recoverable form therefrom. In connection with the transgenic production in mammals, antibodies can be produced in, and recovered from, the milk of goats, cows, or other mammals. *See, e.g.*, U.S.
10 Patent Nos. 5,827,690, 5,756,687, 5,750,172, and 5,741,957.

[0058] The invention contemplates an isolated human antibody or antigen-binding portion thereof that was expressed in a non-human animal and specifically binds to the LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016
15 (02/05), and 170006 (02). In a preferred embodiment, the isolated human antibody or antigen-binding portion thereof is a monoclonal antibody.

[0059] The invention further contemplates the isolated human antibody or antigen-binding portion thereof that is opsonic for *P. aeruginosa* cells from one of strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6
20 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the isolated human antibody or antigen-binding portion thereof facilitates phagocytosis of the *P. aeruginosa* cells.

[0060] The invention also contemplates that the isolated human antibody or antigen-binding portion thereof enhances the immune response to *P. aeruginosa* from one of
25 strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the isolated human antibody or antigen-binding portion thereof facilitates the killing of *P. aeruginosa* cells. In a more preferred embodiment, the isolated human antibody or antigen-binding portion thereof facilitates the killing of *P.*
30 *aeruginosa* cells by delivering an agent that is lethal to the *P. aeruginosa* cells.

[0061] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher

Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), wherein the antibody or antigen-binding portion thereof inhibits *P. aeruginosa* infection.

5 [0062] The invention also contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), wherein the antibody or antigen-binding portion thereof binds to *P. aeruginosa* LPS with a dissociation constant (K_d) of 5×10^{-7} M or less, preferably 5×10^{-7} M to 1×10^{-7} M.

10 In a more preferred embodiment, the antibody or antigen-binding portion thereof binds to *P. aeruginosa* LPS with a K_d of 1×10^{-7} M to 5×10^{-8} M. In a more preferred embodiment, the antibody or antigen-binding portion thereof binds to *P. aeruginosa* LPS with a K_d of 5×10^{-8} M to 1×10^{-8} M.

[0063] The invention contemplates an isolated human antibody or antigen-binding

15 portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and has a half-life *in vivo* of one hour or more. In a preferred embodiment, the antibody or antigen-binding portion thereof has a half-life *in vivo* of between one hour and thirty days. In a more

20 preferred embodiment, the antibody or antigen-binding portion thereof has a half-life *in vivo* of between sixteen and thirty days. In another more preferred embodiment, the antibody or antigen-binding portion thereof has a half-life *in vivo* of between one hour and fifteen days.

[0064] The isolated human antibody or antigen-binding portion thereof that

25 specifically binds to *P. aeruginosa* LPS of the invention may be immunoglobulin G (IgG), IgM, IgE, IgA and IgD. In a preferred embodiment, the IgG may be an IgG1, IgG2, IgG3 or IgG4 subtype. In some preferred embodiments, the IgG is the IgG2 subtype.

[0065] The invention contemplates an isolated human antibody or antigen-binding

30 portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and is labeled. In a

preferred embodiment, the label is a radiolabel, an enzyme label, a fluorescent label, a toxin, a magnetic agent, a second antibody, an affinity label, an epitope tag, an antibiotic, a complement protein or a cytokine.

[0066] The invention contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and comprises a kappa light chain and framework sequences thereof. In preferred embodiments, the framework sequences of the kappa light chain are encoded by a human gene selected from the group consisting of: V κ 2/A19/A3; V κ 1/A30; V κ 4/B3; V κ 3/A27; V κ 3/L2; V κ 1/A30; V κ 3/L2,L16; and V κ 1/A30. In a preferred embodiment, the kappa light chain comprises between one and fifteen changes from a kappa light chain encoded by the germline sequence of one of these genes. In a more preferred embodiment, the kappa light chain comprises no more than six amino acid changes from a kappa light chain encoded by the germline sequence of one of these genes. In a more preferred embodiment, the kappa light chain comprises no more than three amino acid changes from a kappa light chain encoded by the germline sequence of one of these genes.

[0067] The invention contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and comprises a kappa light chain comprising an amino acid sequence selected from the group consisting of: SEQ ID NO: 22; SEQ ID NO: 23; SEQ ID NO: 24; SEQ ID NO: 25; SEQ ID NO: 26; SEQ ID NO: 27; SEQ ID NO: 28; SEQ ID NO: 29 and SEQ ID NO: 30. The invention also contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and comprises a kappa light chain comprising at least one of the FR1, CDR1, FR2, CDR2, FR3, CDR3 and J regions sequence from an amino acid sequence selected from the group consisting of: SEQ ID NO: 22; SEQ ID NO: 23; SEQ ID NO: 24; SEQ ID NO: 25; SEQ ID NO: 26; SEQ ID NO: 27; SEQ ID NO: 28; SEQ ID NO: 29 and SEQ ID NO: 30. The

invention also contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to *P. aeruginosa* LPS and comprises a lambda light chain.

5 [0068] The invention contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to *P. aeruginosa* LPS, comprising a heavy chain composed of variable (V), diversity (D), and Joining (J) regions and composed of framework sequences thereof. In a preferred embodiment, the variable region of the heavy chain is encoded by a human gene selected from the group consisting of: V_H3/V4-04; V_H3/V4-59; V_H3/V3-33; V_H3/V3-15; V_H6/V6-01; and V_H5/V5-51. In
10 another preferred embodiment, the diversity region of the heavy chain is encoded by a human gene selected from the group consisting of: D3-10; D1-26; D3-22; D6-13; and D6-19. In another preferred embodiment, the joining region of the heavy chain is encoded by a human J_H3, J_H4 or J_H6 gene. In a more preferred embodiment, the variable region comprises between one and fifteen amino acid changes from a
15 variable region encoded by the germline gene. In a more preferred embodiment, the heavy chain comprises no more than six amino acid changes from a variable region encoded by the germline gene. In a more preferred embodiment, the heavy chain comprises no more than three amino acid changes from a variable region encoded by the germline gene.

20 [0069] The invention contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and comprises a heavy chain comprising an amino acid sequence selected from the group consisting of: SEQ ID
25 NO: 13; SEQ ID NO: 14; SEQ ID NO: 15; SEQ ID NO: 16; SEQ ID NO: 17; SEQ ID NO: 18; SEQ ID NO: 19; SEQ ID NO: 20; and SEQ ID NO: 21. The invention also contemplates an isolated antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003
30 (02), IATS016 (02/05), and 170006 (02) and comprises a heavy chain comprising at least one of the FR1, CDR1, FR2, CDR2, FR3, CDR3 and J regions sequence from an amino acid sequence selected from the group consisting of: SEQ ID NO: 13; SEQ ID

NO: 14; SEQ ID NO: 15; SEQ ID NO: 16; SEQ ID NO: 17; SEQ ID NO: 18; SEQ ID NO: 19; SEQ ID NO: 20; and SEQ ID NO: 21.

[0070] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and comprises an antigen binding domain chosen from the list consisting of an Fab fragment, an F(ab')₂ fragment, an F_v fragment, a single chain antibody, a humanized antibody, a chimeric antibody or a bispecific antibody.

[0071] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and the antibody is a single chain antibody.

[0072] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and the antibody is a chimeric antibody. In a preferred embodiment, the chimeric antibody comprises framework regions and CDR regions from different human antibodies. In a more preferred embodiment, the chimeric antibody is bispecific. In a more preferred embodiment, the chimeric antibody is bispecific for *P. aeruginosa* LPS and a label selected from the list consisting of a radiolabeled molecule, an enzymatic label, a fluorescent label, a toxin, a magnetic agent, a second antibody, an affinity label, an epitope tag, an antibiotic, a complement protein and a cytokine.

[0073] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and the antibody or portion thereof is derivatized. In a preferred embodiment, the antibody or portion thereof is derivatized with polyethylene glycol, at least one methyl or ethyl group or at least one carbohydrate moiety.

[0074] The invention contemplates a pharmaceutical composition comprising an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and a pharmaceutically acceptable carrier. In certain embodiments, the invention contemplates a pharmaceutical composition comprising more than one isolated human antibody or antigen-binding portion thereof that specifically binds to *P. aeruginosa* LPS and a pharmaceutically acceptable carrier. The invention further contemplates a kit comprising the antibody or antigen-binding portion thereof or antibodies or antigen-binding portions thereof, a pharmaceutically acceptable carrier therefor, and a container. In a preferred embodiment, the kit further comprising instructions for use.

[0075] The invention contemplates a method for treating or preventing *P. aeruginosa* infection, comprising the step of administering a pharmaceutical composition to a patient at risk of being infected with, or currently infected with, *P. aeruginosa*.

[0076] In a preferred embodiment, the human antibody is a monoclonal antibody. In another preferred embodiment, the pharmaceutical composition is administered via an injection, transmucosal, oral, inhalation, ocular, rectal, long acting implantation, liposomes, emulsion, cream, topical or sustained release means. In another preferred embodiment, the antibody is a fusion with a second protein. In a more preferred embodiment the second protein is chosen from the list consisting of a toxic peptide moiety, a complement protein, a radiolabeled protein, a cytokine or an antibiotic protein. In another preferred embodiment, the antibody is labeled with a radiolabel, a toxin, a complement protein, a cytokine or an antibiotic. In another preferred embodiment, the patient is a burn patient, a surgical patient, a prosthesis recipient, a respiratory patient, a cancer patient, a cystic fibrosis patient or an immunocompromised patient. In another preferred embodiment, the pharmaceutical composition further comprises toxins, complement proteins, radiolabeled proteins, cytokines, antibiotics, or any combination thereof.

[0077] The invention contemplates an isolated cell line that produces a human antibody or antigen-binding portion thereof that specifically binds to LPS from one of

P. aeruginosa strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the cell line is a hybridoma.

[0078] The invention contemplates a method of producing an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), comprising:

a) culturing a non-human cell capable of producing the antibody under conditions in which the antibody is produced;

b) isolating the antibody from the cell culture.

[0079] In a preferred embodiment, the method of producing an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) utilizes a hybridoma. In another preferred embodiment, the method utilizes a cell that is transformed with isolated nucleic acid molecules encoding the human antibody or antigen-binding portion thereof and the cell is chosen from the list consisting of a bacterial cell, a yeast cell, an insect cell, an amphibian cell and a mammalian cell. In a more preferred embodiment, the mammalian cell is selected from the list consisting of a human cell, a mouse cell, a rat cell, a dog cell, a monkey cell, a goat cell, a pig cell, a bovine cell and a hamster cell. In a more preferred embodiment, the mammalian cell is selected from the list consisting of a HeLa cell, a NIH 3T3 cell, a CHO cell, a BHK cell, a VERO cell, a CV-1 cell, a NS/0 cell and a COS cell.

[0080] The invention contemplates a nucleic acid molecule isolated from a non-human animal that encodes a human antibody heavy chain or the antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the nucleic acid molecule is derived from a hybridoma that produces the human antibody.

[0081] The invention contemplates a vector comprising a nucleic acid molecule, or fragment thereof, encoding a human antibody heavy chain or antigen-binding portion thereof that specifically binds one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the vector further comprises expression control sequences operably linked to the nucleic acid.

[0082] The invention contemplates a nucleic acid molecule isolated from a non-human animal that encodes a human antibody light chain or the antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the nucleic acid molecule is isolated from a hybridoma that produces the human antibody.

[0083] The invention contemplates a vector comprising a nucleic acid molecule, or fragment thereof, encoding a human antibody light chain or antigen-binding portion thereof that specifically binds to *P. aeruginosa* from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). In a preferred embodiment, the vector further comprises an expression control sequence operably linked to the nucleic acid.

[0084] The invention contemplates an isolated host cell comprising

a) a nucleic acid molecule that was isolated from a non-human animal and encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02); or

b) a vector comprising the nucleic acid molecule.

[0085] The invention contemplates an isolated host cell comprising:

a) a nucleic acid molecule that was isolated from a non-human animal and encodes a heavy chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *P. aeruginosa* strains Fisher

Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02); or

b) a vector comprising the nucleic acid molecule.

[0086] The invention contemplates an isolated host cell comprising:

5 a) a nucleic acid molecule that was isolated from a non-human animal and encodes a heavy chain or the antigen-binding portion thereof and an isolated nucleic acid molecule that encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4
10 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02); or

b) a vector or vectors comprising the nucleic acid molecules.

[0087] In a preferred embodiment, the isolated host cells described above are chosen from the list consisting of hybridoma cells, bacterial cells, yeast cells, insect
15 cells, amphibian cells and mammalian cells. In a more preferred embodiment, the mammalian cells are selected from the list consisting of human cells, mouse cells, rat cells, dog cells, monkey cells, goat cells, pig cells, bovine cells and hamster cells. In a more preferred embodiment, the mammalian cells are selected from the list consisting of HeLa cells, NIH 3T3 cells, CHO cells, BHK cells, VERO cells, CV-1
20 cells, NS/0 cells and COS cells.

[0088] The invention contemplates a method of recombinantly producing the heavy chain or the antigen-binding portion thereof, the light chain or the antigen-binding portion thereof, or both the light chain and heavy chain or antigen-binding portions thereof, of a human antibody that was identified from a non-human animal and
25 specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), comprising the step of cultivating the host cells described above under conditions in which the nucleic acid molecules are expressed.

30 **[0089]** The invention contemplates an isolated human antibody heavy chain or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4

(01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), encoded by any of the nucleic acid molecules encoding the heavy chain described above, or isolated from any of the host cells described above. In a preferred embodiment, the isolated human antibody heavy chain or antigen-binding portion thereof comprises between one to ten amino acid substitutions that increase the serum half-life of the antibody.

[0090] The invention contemplates an isolated human antibody light chain or antigen-binding portion thereof that specifically binds to LPS from one of *P.*

aeruginosa strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), encoded by any of the nucleic acid molecules encoding the heavy chain described above, or isolated from any of the host cells described above. In a preferred embodiment, the isolated human antibody light chain or antigen-binding portion thereof comprises between one to ten amino acid substitutions that increase the serum half-life of the antibody.

[0091] The invention contemplates a non-human transgenic animal comprising any of the nucleic acid molecules described above. In a preferred embodiment, the non-human transgenic animal expresses the nucleic acid molecule or molecules. In a more preferred embodiment, the non-human transgenic animal comprises an isolated nucleic acid molecule that encodes a heavy chain or the antigen-binding portion thereof and an isolated nucleic acid molecule that encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), and the non-human animal expresses both nucleic acid molecules. In a more preferred embodiment, the non-human animal is selected from the list consisting of a mouse, a rat, a hamster, a cow, a sheep, a primate, a horse and a pig. In a more preferred embodiment, a human antibody resulting from expression of the isolated nucleic acid molecules or portions thereof is expressed on the surface of cells derived from the animal's B lymphocytic cells or progeny thereof. In another preferred embodiment, the human antibody resulting from expression of the isolated nucleic

acid molecules or a portion thereof is secreted into the lymph, blood, milk, saliva, or ascites of the animal.

5 [0092] The invention contemplates a fusion protein comprising the an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) and a second polypeptide sequence. In a preferred embodiment, the second polypeptide sequence is chosen from the list consisting of an epitope tag, an affinity tag, a toxic polypeptide, an antibiotic, an enzyme, a second antibody
10 sequence, a complement protein, and a cytokine.

[0093] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), wherein the heavy chain
15 isotype of the antibody is mu, gamma, delta, epsilon or alpha.

[0094] The invention contemplates an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), wherein the antibody or
20 antigen-binding portion thereof is produced by a process comprising the steps of:

a) immunizing a non-human animal comprising a human immunoglobulin locus with an antigen selected from the group consisting of an isolated *P. aeruginosa* LPS preparation, a virulent *P. aeruginosa* cell preparation, an attenuated *P. aeruginosa* cell preparation, and a killed *P. aeruginosa* cell preparation,
25 wherein the *P. aeruginosa* is one of strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02);

b) allowing the non-human animal to mount an immune response to the antigen; and

30 c) isolating the antibody from the non-human animal.

[0095] The invention contemplates an isolated human antibody or antigen-binding portion thereof isolated from an animal or cell that was free of contaminating human

biomaterials. In a preferred embodiment, the biomaterials are viruses, enzymes, hormones, cytokines, receptors, receptor ligands, immunoglobulins, complement, nuclear proteins, and cytoplasmic signaling proteins. In a more preferred embodiment, the viruses are Epstein-Barr virus or retroviruses.

5 **[0096]** Pharmaceutical compositions may be manufactured by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes.

10 **[0097]** Pharmaceutical compositions for use in accordance with the present invention thus may be formulated in a conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Proper formulation is dependent upon the route of administration chosen.

15 **[0098]** For injection, the agents of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks's solution, Ringer's solution, or physiological saline buffer. For transmucosal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art. For ocular administration, suspensions in an appropriate saline solution are used as is well known in the art.

20 **[0099]** For oral administration, the compounds can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers well known in the art. Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a patient to be treated. Pharmaceutical
25 preparations for oral use can be obtained as a solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients include fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato
30 starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired,

disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof such as sodium alginate.

[0100] Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures.

Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

[0101] Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the active ingredients in admixture with fillers such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added. All formulations for oral administration should be in dosages suitable for such administration.

[0102] For buccal administration, the compositions may take the form of tablets or lozenges formulated in conventional manner.

[0103] For administration by inhalation, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant, *e.g.*, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, *e.g.*, gelatin, for use in an inhaler or insufflator, may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

[0104] The compounds may be formulated for parenteral administration by injection, *e.g.*, by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, *e.g.*, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions,

solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents.

[0105] Pharmaceutical formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form. Additionally, suspensions
5 of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran.
10 Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

[0106] Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, such as sterile pyrogen-free water, before use.

[0107] The compounds may also be formulated in rectal compositions such as
15 suppositories or retention enemas, *e.g.*, containing conventional suppository bases such as cocoa butter or other glycerides.

[0108] In addition to the formulations described previously, the compounds may also be formulated as a depot preparation. Such long acting formulations may be
20 administered by implantation (for example subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

[0109] A pharmaceutical carrier for the hydrophobic compounds of the invention is a cosolvent system comprising benzyl alcohol, a nonpolar surfactant, a water-miscible organic polymer, and an aqueous phase. The cosolvent system may be the VPD co-solvent system. VPD is a solution of 3% w/v benzyl alcohol, 8% w/v of the nonpolar surfactant polysorbate 80, and 65% w/v polyethylene glycol 300, made up to volume
25 in absolute ethanol. The VPD co-solvent system (VPD:5W) consists of VPD diluted 1:1 with a 5% dextrose in water solution. This co-solvent system dissolves
30 hydrophobic compounds well, and itself produces low toxicity upon systemic

administration. Naturally, the proportions of a co-solvent system may be varied considerably without destroying its solubility and toxicity characteristics.

Furthermore, the identity of the co-solvent components may be varied: for example, other low-toxicity nonpolar surfactants may be used instead of polysorbate 80; the
5 fraction size of polyethylene glycol may be varied; other biocompatible polymers may replace polyethylene glycol, *e.g.*, polyvinyl pyrrolidone; and other sugars or polysaccharides may be substituted for dextrose.

[0110] Alternatively, other delivery systems for hydrophobic pharmaceutical compounds may be employed. Liposomes and emulsions are well known examples
10 of delivery vehicles or carriers for hydrophobic drugs. Certain organic solvents such as dimethylsulfoxide also may be employed, although usually with a greater toxicity.

[0111] Additionally, the compounds may be delivered using a sustained-release system, such as semipermeable matrices of solid hydrophobic polymers containing the therapeutic agent. Various sustained-release materials have been established and
15 are well known by those skilled in the art. Sustained-release capsules may, depending on their chemical nature, release the compounds for a few weeks up to over 100 days.

[0112] Depending on the chemical nature and the biological stability of the therapeutic reagent, additional strategies for protein stabilization may be employed.

[0113] The pharmaceutical compositions also may comprise suitable solid or gel
20 phase carriers or excipients. Examples of such carriers or excipients include but are not limited to calcium carbonate, calcium phosphate, various sugars, starches, cellulose derivatives, gelatin, and polymers such as polyethylene glycols.

[0114] The isolated human antibody or antigen-binding portion thereof that specifically binds to *P. aeruginosa* LPS of the invention may be provided as salts with
25 pharmaceutically compatible counterions. Pharmaceutically compatible salts may be formed with many acids, including but not limited to hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, etc. Salts tend to be more soluble in aqueous or other protonic solvents that are the corresponding free base forms.

[0115] A component of the kits of the present invention comprise instructions for
30 utilizing the compositions of the present invention for prevention or treatment of *P. aeruginosa* infections. Applicant has, for the first time, disclosed herein a method of preventing or treating *P. aeruginosa* infections with an isolated human antibody or

antigen-binding portion thereof that specifically binds to LPS from one of *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). The printed instructions on the kit enable one of skill in the art to utilize the kit
5 for practicing the methods of the present invention.

EXAMPLE 1

Generation of Mice and Hybridomas That Produce Fully Human Antibodies to P. aeruginosa LPS

10 [0116] *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-1 (06), It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02) — all of which were originally clinical isolates — were used for mouse immunizations, mouse protection assays and opsonic assays. Bacteria
15 for mouse challenge assays were freshly plated onto pseudosel agar (BBL, Becton Dickinson, Sparks, MD), then were incubated at 37°C, and cells from a single colony were inoculated into LB broth and incubated at 37°C in a shaking water bath to a concentration of 5×10^8 cfu/ml. Bacteria were centrifuged at 10,000 rpm for 10 minutes, resuspended and washed in chilled phosphate buffered saline (PBS) and
20 diluted as needed. Bacteria for immunization experiments were grown as above, heat-killed at 60°C for one hour and stored at 4°C until use.

[0117] The high molecular weight polysaccharide (high MW PS) portion of the LPS O-specific side chains from *P. aeruginosa* strains Fisher Devlin (International Serogroups) It-1 (06), It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05),
25 170003 (02), IATS016 (02/05), and 170006 (02) were made as described previously, and were lyophilized for storage. See Hatano et al. *Infect. Immun.* 62:3608-3616 (1994). These high MW PS were used to block binding of Mab to bacteria immobilized on microtiter plates for enzyme-linked immunosorbent assays (ELISA) as described in Example 3.

30 [0118] Mice that were transgenic for human heavy and light Ig were bred and maintained by Abgenix Inc., Fremont, CA. The strain of XenomouseTM animals used was XMG2, which is an Ig-inactivated mouse reconstituted with a YAC containing cointegrated human heavy and light chain transgenes as previously described. See

Mendez et al. *Nature Genet.* **15**:146-56 (1997). Mice were housed in micro-isolator cages in a pathogen-free facility after shipping, and food and water were autoclaved prior to use. Mice were immunized with 10^7 heat-killed *P. aeruginosa* of the various strains intraperitoneally (ip; 10^7 bacteria in PBS) with Freund's complete adjuvant
5 (Sigma, St. Louis MO) for the first injection and Freund's incomplete adjuvant two weeks later and for the remainder of the four weekly injections. Mouse sera was obtained after four to six weeks of immunizations via tail vein bleed and were screened for anti-O-specific side chain antibodies by ELISA as described below in Example 3. Seropositive mice were boosted by intravenous (iv) injection of 1×10^7 cfu
10 of heat-killed *P. aeruginosa* bacteria in sterile PBS without any adjuvant four days before splenectomy and fusion were performed.

[0119] Hybridomas were generated by fusing spleen and/or lymph node cells from immunized, seropositive XenomouseTM animals with the nonsecreting sp2/0 myeloma cell line, as described previously. See Mendez et al. *Nature Genet.* **15**:146-156
15 (1997); Schreiber et al. *J. Immunol.* **146**:188-193 (1991). Supernatants from hybridomas were screened for production of human anti-*P. aeruginosa* LPS Mabs using the ELISA procedure described below in Example 3. Hybridomas found to be secreting IgG anti-LPS antibodies were then cloned three times by limiting dilution. One IgG2-secreting clone was chosen for each *P. aeruginosa* strain and designated
20 according to the strain against which they were raised as follows: anti-It-2; anti-It-3; anti-It-4; anti-It-5; anti-It-6; anti-PA01; anti-170003; anti-IATS016; and anti-170006.

EXAMPLE 2

25 *Characterization and Usage of Variable Region Genes From Transgenic Mouse-derived Anti-LPS Antibody*

[0120] Human heavy-chain and light-chain variable (V) regions were cloned into a T7-promoter driven vector by amplifying the variable regions. The heavy-chain variable (V_H) region and the light-chain variable (V_L) region primers used are
30 included in Table 1. These primers were synthesized by Integrated DNA Technologies (Coralville, IA). PCR was run at 94°C, 60sec; 50°C, 60 sec; 72°C, 120 sec for 35 cycles. The PCR products were run on a 2% Tris-acetate EDTA agarose gel and the bands (423 bp V_K , 441bp V_H) were isolated using the QIAquick[®] gel-

extraction kit (Qiagen, Inc., Valencia, CA). The cDNA was cloned into the pT7Blue vector and transformed into *Escherichia coli* using the Perfectly Blunt® Cloning Kit from Novagen (EMD Biosciences, Inc.). Positive clones were grown overnight in LB-ampicillin broth. The DNA was extracted from the bacteria and purified by Qiagen® Miniprep. DNA samples were sent to Cleveland Genomics for sequencing. V region sequences were compared and classified using V Base to determine gene usage for each *P. aeruginosa* lipopolysaccharide serotype.

Table 1. Primers used to amplify heavy-chain and light-chain V regions.

Primer	Sequence	SEQ ID NO:
V _H 3'	5'-CCC AAG CTT TTC GGC GAA GTA GTC CTT GAC CAG GCA GCC CAG-3'	1
con IgG2	5'-GCA CTC ACT AGT ACA TTT GCG CTC AAC-3'	2
V _H A	5'-GGG AAT TCA TGG ACT GGA CCT GGA GGR TYC TCT KC-3'	3
V _H B	5'-GGG AAT TCA TGG AGY TTG GGC TGA SCT GGS TTT YT-3'	4
V _H C	5'-GGG AAT TCA TGR AMM WAC TKT GKW SCW YSC TYC TG-3'	5
V _H 1a	5'-GAG GTR CAG YTG CTC GAG TCT GGR G-3'	6
V _H 1b	5'-CAG ACK CAG YTG CTC GAG TCT GGG RGC-3'	7
V _H 2	5'-CAG GTG CAG CTG CTC GAG TCG GGC-3'	8
V _H 3	5'-GAG GTG CAG CTG CTC GAG TCT GG-3'	9
V _H 4	5'-CAG GWG CAG CTG CTC GAG TCK GGG-3'	10
V _L 3'	5'-CCC AAG CTT CAT CAG ATG GCG GGA AGA-3'	11
V _L 1	5'-GGG AAT TCA TGG ACA TGR RRD YCC HVG YKC ASC TT-3'	12

10

[0121] Immunization of the transgenic mice with heat-killed *P. aeruginosa* resulted in the production of IgM and IgG2 human antibodies directed to the LPS O-side chain of the serotype used for immunization, consistent with the constant region reconstitution of this mouse (data not shown). Only IgG2 antibodies were chosen for further characterization. Variable region genes from hybridomas obtained from fusion of spleen cells from *P. aeruginosa*-immunized transgenic mice with the non-secreting sp2/0 cell line were cloned and sequenced in order to determine variable region gene

15

usage. The deduced amino acid sequences of the V regions of the heavy and light chains of these monoclonal antibodies are shown in Tables 2 and 3. The amino acid sequences in Tables 2 and 3 constitute continuous sequences, but are separated by the symbol “—” only to indicate the junctions of the corresponding FR1, CDR1, FR2, CDR2, FR3, CDR3, and J regions, respectively.

Table 2. Amino acid sequences of the V regions of the heavy chains.

Mab	Heavy Chain Sequence	SEQ ID NO:
Anti-It-2	QVQLQESGPGLVKPSSETLSLTCTVS—GGSISSYYWS— WIRQPAGKGLEWIG—RIYTSNTNYKPSLKS— RVTMSVDTSKNQFSLKLSSVTAADTAVYYCAR— EVMVRGVTFDY—WGQGTLLTVSSA	13
Anti-It-3	QVQLQESGPGLVKPSSETLSLTCTVS—GGSVSDYYWS— WIRQPPGKGLEWIG—YIYYSGSTNYPNPSLKS— RVTISVDTSKNQFSLKLSSVTAADTAVYYCAR— DGSVPPGIY—WGQGTLLTVSSA	14
Anti-It-4	QVQLVESGGGVVQPGRSLRLSCAAS—GFTFRYGMH— WVRQAPGKGLEWVA—VIWYDGNKKYHAESVKG— RFTISRDN SKNTLYLQMNSLRAEDTAVYYCAR— GGFGELPHLYGMDV—WGQGTTTVTVSSA	15
Anti-It-5	EVQLVESGGGLVKPGGSLRLSCAVS— GFTFSNAWMS—WVRQTPGKGLEWVG— RIKSKTDGGTIDYAAPVKG— RFTISRDDSKNTLYLQMNSLKTEDTAVYYCTK— FYYGSGSYGY—WGQGTLLTVTVSSA	16
Anti-It-6	QVQLQQSGPGLVKPSQTLSTCAIS— GDSVSSNSAAWN—WIRQSPSRGLEWLG— RTYYRSKWYNDYAVSVKS— RITINPDTSKNQFSLQLNSVTPEDTAVYYCAR— GYYYGMDV—WGQGTTTVTVSSA	17
Anti-170003	EVQLVESGGGLVKPGGSLRLSCAAS— GFTFSNAWMS—WVRQAPGKGLEWVG— RIKSKTDGGTTDYAAPVKG— RFTISRDDSKNTLYLQMNSLKTEDTAVYYCTT— YYDSSGYYYY—WGQGTLLTVTVSSA	18
Anti-170006	EVQLVQSGAEVKKPGESLKISCKGF—GYSFASYWIG— WVRQMPGKGLEWMG—NIYPGDSYTIYSPSFQG— QVAISADKSISTAYLQWNSLKASDTAMYYCAR— RGFSGRSYDAFEI—WGQGTMTVTVLA	19

Mab	Heavy Chain Sequence	SEQ ID NO:
Anti-Pa01	QVHLQESGPGLVKPSETLSLTCTVS—GGSTITNFIYWS— WIRQSAGKGLEWIG—RIYISGTTNYPNPSLKS— RVTMSLDTSKNQFSLKLSSVTAADTAVYYCAR— GGYSIGWYRDLGSFDI—WGQGTMTVTVSSA	20
Anti-IATS016	QVQLQESGPGLVKPSESLSLTCTVS—GGSVSSYYWS— WIRQPAGKGLEWIG—LIYTSGSTNYPNPSLKS— RVTMSVDTSKNQFSLKLSSVTAADSAVYYCAR— IAAAGTDY—WGQGTMLTVTVSSA	21

Table 3. Amino acid sequences of the V regions of the light chains.

Mab	Heavy Chain Sequence	SEQ ID NO:
Anti-It-2	DIVMTQSPSLPVTTPGEPASISC— RSSQSLLFSNEYNFLD—WFLQKPGQSPQLLIY— LGSNRAS— GVPDRFSGSGSGTDFTLKISRVEAEDVGVYYC— MQALQIPRT—FGQGTKVEIKR	22
Anti-It-3	DIQMTQSPSSLSASVGDRTITC— RASQGIRNVLV—WYQQKPGKAPKRLIY— AASSLQS— GVPSRFSGSGSGTEFTLTISSLQPEDFATYYC— LQHNSYPWT—FGQGTKVEIKR	23
Anti-It-4	DIVMTQSPDSLAVSLGERATINC— KSSQNILYNSNNNNYLA—WFQQKPRQPPKLLIY— WASTRES— GVPDRFSGSGSGTDFTLTINSLQAEDVAVYYC— QQYYSAPLT—FGGGTKVEIKR	24
Anti-It-5	EIVLTQSPGTLSLSPGERATLSC—RTSQSVSSIYLA— WYQQKPGQAPRLIY—GASNRAT— GIPDRFSGSGFGTDFTLTISRLEPEDFAVYYC— QQYGRSPLT—FGGGTKVEIKR	25
Anti-It-6	ERVMTQSPATLSVSPGERATLSC— RASQSVSSNLA—WYQQKPGQAPRLIY— GASTRAT— GIPARFSGSGSGTEFTLTISSLQSEDFAVYYC— QQYYHWLT—FGGGTKVEIKR	26

Anti-170003	DIQMTQSPSSLSASVGDRVITC— RASQGIRNDLG—WYQQKPGKAPKRLIY— AASSLQS— GVPSRFSGSGSGTEFTLTISSLQPEDFATYYC— LQYNSYPPT—FGQGTKVEIKR	27
Anti-170006	EIVMMQSPGPLSVSPGERAILSC RASQNVNINLA— WYQQKPGQAPRLIY—GASTRAT— GIPARFSGSGSGTEFTFTISSLQSEDFAVYYC— QQYKNWPLT—FGGGTKVEIKR	28
Anti-Pa01	DIVMTQSPDSLAVSLGERATINC— KSSQNILYSSNNKNYLA—WYQQKPGQPPKLLIY— WASTRES— GVPDRFSGSGSGTDFTLTISSLQAEDVAVYFC— QQYYNIRT—FGQGTKVEIKR	29
Anti-IATS016	DIQMTQSPSSLSASVGDRVITC— RASQDIRNDLG—WYQQKPGKAPKRLIY— AASSLQS— GVPSRFSGSGSGTEFTLTISSLQPEDFATYYC— LQYKSY PWT—FGQGTKVEIKR	30

[0122] The light-chain gene segments V_K2/A2 and J_K1 were used, as previously reported as commonly used in humans after PS vaccine immunization. See Chung et al. *Infect. Immun.* **63**:4219-4233 (1995). However, many other light-chain gene segments were also used as indicated in Table 4. Three of the new protective IgG2 anti-LPS monoclonal antibodies utilized genes from the V_H3 gene family (anti-It-4, It-5 and 170003) but a variety of other V region genes were utilized by other human anti-LPS O-side chain Mab including V_H4, 5 and 6 (Table 4).

Table 4. V_H and V_K gene usage by human Mabs against LPS O-side chain of *P. aeruginosa* made in transgenic XenoMouse™ mice.

<i>P. aeruginosa</i> LPS O-side chain serotype	Mab isotype	V _H region family	V _K region family
O6ad*	IgG2	V _H 3/V3-33 J _H 4	V _K 2/A2 J _K 1
It-2	IgG2	V _H 4/V4-04 J _H 4	V _K 2/A19/A3 J _K 1
It-3	IgG2	V _H 4/V4-59 J _H 4	V _K 1/A30 J _K 1
It-4	IgG2	V _H 3/V3-33 J _H 6	V _K 4/B3 J _K 4
It-5	IgG2	V _H 3/V3-15 J _H 4	V _K 3/A27 J _K 4
It-6	IgG2	V _H 6/V6-01 J _H 6	V _K 3/L2 J _K 4
170003	IgG2	V _H 3/V3-15 J _H 4	V _K 1/A30 J _K 1
170006	IgG2	V _H 5/V5-51 J _H 3	V _K 3/L2,L16 J _K 4
Pa01	IgG2	V _H 4/V4-04 J _H 3	V _K 3/A27 J _K 4
IATS016	IgG2	V _H 4/V4-04 J _H 4	V _K 1/A30 J _K 1

* This antibody was described in WO 02/20619, published March 14, 2002.

EXAMPLE 3

Detection of Anti-P. aeruginosa LPS antibodies

[0123] Enzyme-linked immunosorbent assay (ELISA) was used to detect antibodies
5 to the various serotypes of *P. aeruginosa* LPS O-side chain in sera of immunized
mice and in hybridoma supernatants as we have previously described. See Schreiber
et al. *J. Immunol.* 146:188-93 (1991). Briefly, 96-well microtiter polystyrene plates
(NUNC, Denmark) were coated with 100 μ l per well of 2 μ g/ml of purified high MW
PS or 1×10^7 cfu/well heat-killed *P. aeruginosa* overnight at 4°C, washed, and blocked
10 with 200 μ l/well of 1% bovine serum albumin (BSA; Sigma-Aldrich, St. Louis, MO)
in PBS and .05 % Tween 20 (Amresco, Solon, OH). Plates were washed and
incubated overnight with serial dilutions of Mab or sera in 1% BSA in PBS. Plates
were washed, and bound antibodies were detected by adding isotype-specific alkaline
phosphatase-conjugated mouse-anti-human polyclonal antibodies (Southern
15 Biotechnology Associates, Birmingham, AL). Plates were developed with 100 μ l/well
of p-nitrophenyl phosphate (PNPP, Sigma-Aldrich) chromogenic substrate in DEA
buffer. Optical densities were measured at 415nm with a microplate reader (Biorad,
Hercules, CA).

[0124] Blocking assays to determine the specificity of Mabs were performed in an
20 identical fashion as above except that soluble *P. aeruginosa* high MW PS or control
PS of different concentrations was added to the the Mab prior to addition to heat-
killed-bacteria-coated 96-well ELISA plates.

[0125] The IgG2 human Mabs produced in the transgenic mouse bound to the O-
side chain of *P. aeruginosa* of nine different strains. Blocking assays revealed over
25 90% reduction in binding of the Mab to heat-killed bacteria after preincubation of the
Mab with the serogroup-specific purified LPS O-side chain, compared to less than
10% inhibition with the control PS (PS from a non-homologous serotype; Fig. 1).
Cross-reaction of Mab, binding with LPS O-side chains from other *P. aeruginosa*
strains occurred, but the observed cross-reaction was always serogroup specific
30 (Table 5).

Table 5. Specificity of human Mabs against *P. aeruginosa* serogroups.

Serotype of <i>P. aeruginosa</i>	Anti-It-1 (O6)*	Anti-It-2 (O11)	Anti-It-3 (O2)	Anti-It-4 (O1)	Anti-It-5 (O10)	Anti-It-6 (O7)	Anti-Pao-1 (O2/O5)	Anti-170003 (O2)	Anti-170006 (O2)	Anti-IATSO-16 (O2)
It-1 (O6)	+++	—	—	—	—	—	—	—	—	—
It-2 (O11)	—	+++	—	—	—	—	—	—	—	—
It-3 (O2)	—	—	+++	—	—	—	—	—	—	+++
It-4 (O1)	—	—	—	+++	—	—	—	—	—	—
It-5 (O10)	—	—	—	—	+++	—	—	—	—	—
It-6 (O7)	—	—	—	—	—	+++	—	—	—	—
It-7 (O2)	—	—	—	—	—	—	+++	—	+++	—
Pao-1 (O2/O5)	—	—	—	—	—	—	+++	—	—	—
170003 (O2)	—	—	—	—	—	—	—	+++	—	—
170006 (O2)	—	—	—	—	—	—	—	—	+++	—
170007 (O2)	—	—	—	—	—	—	—	—	+++	—
IATSO-16 (O2)	—	—	•+++	—	—	—	—	—	—	+++
O6ab	—	—	—	—	—	—	—	—	—	—
O6ac	—	—	—	—	—	—	—	—	—	—
O6ad	+++	—	—	—	—	—	—	—	—	—

* This antibody was described in WO 02/20619, published March 14, 2002.

EXAMPLE 4

Anti-P. aeruginosa LPS Antibody Opsonization Promotes Complement-Dependent Phagocytosis

[0126] The ability of the human monoclonal antibodies to opsonize homologous serotypes of *P. aeruginosa* for uptake by human polymorphonuclear leukocytes (PMN) was measured in a bacterial killing assay as previously described. See Hemachandra et al. *Infect. Immun.* **69**:2223-2229 (2001); Schreiber et al. *J. Infect. Dis.* **167**: 221-226 (1993); and Schreiber et al. *J. Immunol.* **146**:188-193 (1991). Briefly, the killing assay reaction mixture contained 0.1 ml of 1×10^5 cfu/ml of live *P. aeruginosa* in RPMI medium with 10% fetal bovine serum (FBS; endotoxin free, Gibco, Grand Island NY), 0.1 ml of 1×10^7 cells/ml of human PMN (obtained from adult volunteers via venipuncture) in RPMI with 10% FBS, 0.1 ml of different concentrations of human Mab to *P. aeruginosa* in RPMI with 10% FBS, 0.1 ml of 1:15 dilution of human serum from an agammaglobulinemic patient in RPMI with 10% FBS. Controls included human IgG2 Mab of a non-homologous serotype, and a reaction mixture in which PMN were omitted, one in which complement was omitted, and one in which antibody was omitted and replaced with RPMI. After incubation at 37°C with shaking at 100 rpm for 90 minutes, bacteria were diluted and then plated for bacterial enumeration.

[0127] We tested all nine of the new antibodies for their ability to opsonized homologous strains of *P. aeruginosa* (Figure 2). Antibody alone was a mediocre opsonin conforming to our previous findings that Fc γ receptor stimulation without complement receptor stimulation is not optimal for phagocytosis of *P. aeruginosa* by human PMN. See Berger et al. *Pediatr. Res.* **35**:68-77 (1994). Complement alone yielded some increased uptake of labeled bacteria by PMN, but the phagocytosis was greatly enhanced with antibody and complement together, as predicted when both Fc γ and complement receptors are stimulated together in human PMN (data not shown; Berger et al. *supra*). Interestingly, one Mab against It-5 *P. aeruginosa* was non-opsonic in this assay despite specific binding to It-5 bacteria and inhibition of this binding by purified LPS O-side chain. This antibody, however, was protective against fatal sepsis with It-5 bacteria (see below).

EXAMPLE 5

Protection of neutropenic mice from fatal P. aeruginosa sepsis

[0128] In order to determine whether the *in vitro* specificity and opsonic ability of the monoclonal antibodies translated to *in vivo* protective efficacy, the protective efficacy of the human Mab against sepsis caused by homologous serotypes of *P. aeruginosa* was measured in the neutropenic mouse model as we have described previously. See Pier et al. *Infect. Immun.* **57**:174-179 (1989) and Schreiber et al. *J. Immunol.* **146**:188-193 (1991). Female, six week-old BALB/c ByJ mice (Jackson Laboratories, Bar Harbor, ME) were maintained in a pathogen-free, pseudomonas-free environment in which water, bedding, and food were autoclaved prior to use. Neutropenia was established by administering 3 mg of cyclophosphamide (Cytoxan[®], Bristol-Myers Squibb, Princeton, NJ) intraperitoneally (ip) to each mouse on days 1, 3, and 5. On day 5, the cyclophosphamide was administered at time 0 hours, and 2 hours later 25 or 50 mg of antibody was administered ip, followed by 10³ cfu of live *P. aeruginosa* two hours later. Negative control mice received PBS ip since we had previously shown that control mice receiving irrelevant Mab or saline in this model had the same death rates. See Hemachandra et al. *Infect. Immun.* **69**:2223-2229 (2001). Mice were observed daily thereafter for 7 days since all mortality uniformly occurred prior to this endpoint. Cumulative mortality was the outcome measured, but mice that were unable to move were euthanized prior to the 7-day end point since observation indicated that 100% of these mice subsequently died. 5 mice were used for each group.

[0129] Mice receiving saline injection and then challenged with *P. aeruginosa* sustained high mortality, most dying within 48 hours after challenge, consistent with previous descriptions of mortality in non-immune mice in this model. In contrast, those mice receiving the human Mabs derived from the XenoMouse[™] animals were strongly protected from mortality (Figure 3).

[0130] Throughout this specification and claims, the word “comprise,” or variations such as “comprises” or “comprising” will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers.

[0131] All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and
5 example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

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BIOLOGICAL DEPOSITS

[0132] Hybridoma cell lines producing Mabs that recognize the LPS of strains Pa01, It-5, and It-6 were deposited in accordance with the provisions of the Budapest Treaty at the American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, VA 20110-2209, USA on August 6, 2003. They were assigned the following deposit
15 designations and accession numbers:

Anti-Pa01 Hybridoma	PTA-5384	HB _____
Anti-It-5 Hybridoma	PTA-5385	HB _____
Anti It-6 Hybridoma	PTA-5386	HB _____

What is Claimed is:

1. An isolated human antibody or antigen-binding portion thereof that was expressed in a non-human animal and specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02).
2. The isolated human antibody or antigen-binding portion thereof according to claim 1, wherein said human antibody is a monoclonal antibody.
3. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein the antibody or portion thereof is opsonic for *Pseudomonas aeruginosa* cells.
4. The isolated human antibody or antigen-binding portion thereof according to claim 3 wherein said antibody or portion thereof facilitates phagocytosis of said *Pseudomonas aeruginosa* cells.
5. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein the antibody or portion thereof enhances the immune response to *Pseudomonas aeruginosa*.
6. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein the antibody or portion thereof facilitates the killing of *Pseudomonas aeruginosa* cells.
7. The isolated human antibody or antigen-binding portion thereof according to claim 6, wherein said facilitating the killing of *Pseudomonas aeruginosa* cells comprises delivery of an agent lethal to said *Pseudomonas aeruginosa* cells.

8. The isolated human antibody or antigen-binding portion thereof according to claim 6, wherein said facilitating the killing of *Pseudomonas aeruginosa* cells comprises enhancing the immune response to *Pseudomonas aeruginosa*.
9. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or antigen-binding portion thereof inhibits *Pseudomonas aeruginosa* infection.
10. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or antigen-binding portion thereof binds to *Pseudomonas aeruginosa* LPS with a K_d of 5×10^{-7} M to 1×10^{-7} M.
11. The isolated human antibody or antigen-binding portion thereof according to claim 10, wherein said antibody or antigen-binding portion thereof binds to *Pseudomonas aeruginosa* LPS with a K_d of 1×10^{-7} to 5×10^{-8} M.
12. The isolated human antibody or antigen-binding portion thereof according to claim 10, wherein said antibody or antigen-binding portion thereof binds to *Pseudomonas aeruginosa* LPS with a K_d of 5×10^{-8} M to 1×10^{-8} M.
13. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or antigen-binding portion thereof has a half-life *in vivo* of between one hour and thirty days.
14. The isolated human antibody or antigen-binding portion thereof according to claim 13, wherein said antibody or antigen-binding portion thereof has a half-life *in vivo* of between sixteen and thirty days.
15. The isolated human antibody or antigen-binding portion thereof according to claim 13, wherein said antibody or antigen-binding portion thereof has a half-life *in vivo* of between one hour and fifteen days.

16. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody is or is derived from an immunoglobulin molecule having a heavy chain isotype chosen from the list consisting of immunoglobulin G (IgG), IgM, IgE, IgA and IgD.
17. The isolated human antibody according to claim 16, wherein said IgG is a subtype selected from the list consisting of IgG1, IgG2, IgG3 and IgG4.
18. The isolated human antibody according to claim 17, wherein said IgG is the IgG2 subtype.
19. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or portion thereof is labeled.
20. The isolated human antibody or antigen-binding portion thereof according to claim 19, wherein said label is selected from the list consisting of a radiolabel, an enzyme label, a fluorescent label, a toxin, a magnetic agent, a second antibody, an affinity label, an epitope tag, an antibiotic, a complement protein and a cytokine.
21. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, comprising a kappa light chain and framework sequences thereof.
22. The isolated human antibody or antigen-binding portion thereof according to claim 21, wherein said framework sequences of said kappa light chain are encoded by a human gene selected from the group consisting of: V κ 2/A19/A3; V κ 1/A30; V κ 4/B3; V κ 3/A27; V κ 3/L2; V κ 1/A30; V κ 3/L2,L16; and V κ 1/A30.
23. The isolated human antibody or antigen-binding portion thereof according to claim 21, wherein said kappa light chain comprises between one and fifteen amino acid changes from a kappa light chain encoded by the germline sequence of a human gene selected from the group consisting of: V κ 2/A19/A3; V κ 1/A30; V κ 4/B3; V κ 3/A27; V κ 3/L2; V κ 1/A30; V κ 3/L2,L16; and V κ 1/A30.

24. The isolated human antibody or antigen-binding portion thereof according to claim 21, wherein said kappa light chain comprises no more than six amino acid changes from a kappa light chain encoded by the germline sequence of a human gene selected from the group consisting of: V κ 2/A19/A3; V κ 1/A30; V κ 4/B3; V κ 3/A27; V κ 3/L2; V κ 1/A30; V κ 3/L2,L16; and V κ 1/A30.

25. The isolated human antibody or antigen-binding portion thereof according to claim 24, wherein said kappa light chain comprises no more than three amino acid changes from a kappa light chain encoded by the germline sequence of a human gene selected from the group consisting of: V κ 2/A19/A3; V κ 1/A30; V κ 4/B3; V κ 3/A27; V κ 3/L2; V κ 1/A30; V κ 3/L2,L16; and V κ 1/A30.

26. The isolated human antibody or antigen-binding portion thereof according to claim 21, wherein said kappa light chain comprises an amino acid sequence selected from the group consisting of: SEQ ID NO: 22; SEQ ID NO: 23; SEQ ID NO: 24; SEQ ID NO: 25; SEQ ID NO: 26; SEQ ID NO: 27; SEQ ID NO: 28; SEQ ID NO: 29; and SEQ ID NO: 30.

27. The isolated human antibody or antigen-binding portion thereof according to claim 21, wherein said kappa light chain comprises at least one of the FR1, CDR1, FR2, CDR2, FR3, CDR3 and J regions sequence from an amino acid sequence selected from the group consisting of: SEQ ID NO: 22; SEQ ID NO: 23; SEQ ID NO: 24; SEQ ID NO: 25; SEQ ID NO: 26; SEQ ID NO: 27; SEQ ID NO: 28; SEQ ID NO: 29 and SEQ ID NO: 30.

28. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, comprising a lambda light chain.

29. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, comprising a heavy chain composed of variable (V), diversity (D), and Joining (J) regions and composed of framework sequences thereof.

30. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said variable region of said heavy chain is encoded by a human gene selected from the group consisting of: V_H3/V4-04; V_H3/V4-59; V_H3/V3-33; V_H3/V3-15; V_H6/V6-01; and V_H5/V5-51.
31. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said diversity region of said heavy chain is encoded by a human gene selected from the group consisting of: D3-10; D1-26; D3-22; D6-13; and D6-19.
32. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said joining region of said heavy chain is encoded by a human gene selected from the group consisting of: J_H3, J_H4 and J_H6.
33. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said variable region comprises between one and fifteen amino acid changes from a variable region encoded by the germline sequence of a human gene selected from the group consisting of: V_H3/V4-04; V_H3/V4-59; V_H3/V3-33; V_H3/V3-15; V_H6/V6-01; and V_H5/V5-51.
34. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said heavy chain comprises no more than six amino acid changes from a variable region encoded by the germline sequence of a human gene selected from the group consisting of: V_H3/V4-04; V_H3/V4-59; V_H3/V3-33; V_H3/V3-15; V_H6/V6-01; and V_H5/V5-51.
35. The isolated human antibody or antigen-binding portion thereof according to claim 29, wherein said heavy chain comprises no more than three amino acid changes from a variable region encoded by the germline sequence of a human gene selected from the group consisting of: V_H3/V4-04; V_H3/V4-59; V_H3/V3-33; V_H3/V3-15; V_H6/V6-01; and V_H5/V5-51.

36. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said heavy chain comprises an amino acid sequence selected from the group consisting of: SEQ ID NO: 13; SEQ ID NO: 14; SEQ ID NO: 15; SEQ ID NO: 16; SEQ ID NO: 17; SEQ ID NO: 18; SEQ ID NO: 19; SEQ ID NO: 20; and SEQ ID NO: 21.

37. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2 wherein said heavy chain comprises at least one of the FR1, CDR1, FR2, CDR2, FR3, CDR3 and J regions sequence from an amino acid sequence selected from the group consisting of: SEQ ID NO: 13; SEQ ID NO: 14; SEQ ID NO: 15; SEQ ID NO: 16; SEQ ID NO: 17; SEQ ID NO: 18; SEQ ID NO: 19; SEQ ID NO: 20; and SEQ ID NO: 21.

38. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2 comprising an antigen binding domain chosen from the list consisting of an Fab fragment, an F(ab')₂ fragment and an Fv fragment.

39. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody is a single chain antibody.

40. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody is a chimeric antibody.

41. The chimeric antibody according to claim 40, comprising framework regions and CDR regions from different human antibodies.

42. The chimeric antibody according to claim 40, wherein the chimeric antibody is bispecific.

43. The chimeric antibody according to claim 42, wherein the chimeric antibody is bispecific for *Pseudomonas aeruginosa* LPS and a label selected from the list consisting of a radiolabeled molecule, an enzymatic label, a fluorescent label, a toxin,

a magnetic agent, a second antibody, an affinity label, an epitope tag, an antibiotic, a complement protein and a cytokine.

44. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or portion thereof is derivatized.

45. The isolated human antibody or antigen-binding portion thereof according to claim 44, wherein said antibody or portion thereof is derivatized with polyethylene glycol, at least one methyl or ethyl group or at least one carbohydrate moiety.

46. A pharmaceutical composition comprising the antibody or antigen-binding portion thereof according to claim 1 or claim 2 and a pharmaceutically acceptable carrier.

47. A kit comprising the antibody or antigen-binding portion thereof according to claim 1 or claim 2, a pharmaceutically acceptable carrier therefor, and a container.

48. The kit according to claim 47, further comprising instructions for use.

49. A method for treating or preventing *Pseudomonas aeruginosa* infection, comprising the step of administering an isolated human antibody according to claim 1 to a patient at risk of being infected with, or currently infected with, *Pseudomonas aeruginosa*.

50. The method according to claim 49 wherein said human antibody is a monoclonal antibody.

51. The method according to claim 49, wherein said administering is performed via an injection, transmucosal, oral, inhalation, ocular, rectal, long acting implantation, liposomes, emulsion, cream, topical or sustained release means.

52. The method according to claim 49, wherein said antibody is a fusion with a second protein.
53. The method according to claim 49, wherein said antibody is labeled with a radiolabel, a toxin, a complement protein, a cytokine or an antibiotic.
54. The method according to claim 52, wherein said second protein is chosen from the list consisting of a toxic peptide moiety, a complement protein, a radiolabeled protein, a cytokine or an antibiotic protein.
55. The method according to claim 49 wherein said patient is a burn patient, a surgical patient, a prosthesis recipient, a respiratory patient, a cancer patient, a cystic fibrosis patient or an immunocompromised patient.
56. The method according to claim 49, wherein said pharmaceutical composition further comprises toxins, complement proteins, radiolabeled proteins, cytokines, antibiotics, or any combination thereof.
57. An isolated cell line that produces the antibody according to claim 1 or claim 2.
58. The cell line according to claim 57 wherein said cell line is a hybridoma.
59. A method of producing an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), comprising:
- a) culturing a non-human cell capable of producing said antibody under conditions in which the antibody is produced;
 - b) isolating said antibody from said cell culture.
60. The method according to claim 59, wherein said cell is a hybridoma.

61. The method according to claim 59, wherein said cell is transformed with isolated nucleic acid molecules encoding said human antibody or antigen-binding portion thereof and said cell is chosen from the list consisting of a bacterial cell, a yeast cell, an insect cell, an amphibian cell and a mammalian cell.

62. The method according to claim 61, wherein said mammalian cell is selected from the list consisting of a human cell, a mouse cell, a rat cell, a dog cell, a monkey cell, a goat cell, a pig cell, a bovine cell and a hamster cell.

63. The method according to claim 61, wherein said mammalian cell is selected from the list consisting of a HeLa cell, a NIH 3T3 cell, a CHO cell, a BHK cell, a VERO cell, a CV-1 cell, a NS/O cell and a COS cell.

64. A method for making an isolated human antibody or antigen-binding portion thereof that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), IATS016 (02/05), and 170006 (02), comprising:

- a) immunizing a non-human animal having incorporated a human immunoglobulin locus therein with a *Pseudomonas aeruginosa* antigenic composition;
- b) allowing said non-human animal to mount a humoral response to said antigenic composition; and
- c) isolating said human antibody from said non-human animal.

65. A nucleic acid molecule isolated from a non-human animal that encodes a human antibody heavy chain or the antigen-binding portion thereof that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02).

66. The isolated nucleic acid molecule according to claim 65, wherein said nucleic acid molecule is isolated from a hybridoma that produces said human antibody.

67. An isolated nucleic acid molecule, or a fragment thereof, encoding a human antibody heavy chain or antigen-binding portion thereof that specifically binds to *Pseudomonas aeruginosa* LPS comprising a nucleotide sequence encoding an amino acid sequence selected from the group consisting of: SEQ ID NO: 13; SEQ ID NO: 14; SEQ ID NO: 15; SEQ ID NO: 16; SEQ ID NO: 17; SEQ ID NO: 18; SEQ ID NO: 19; SEQ ID NO: 20; and SEQ ID NO: 21.

68. The isolated nucleic acid molecule of claim 67 wherein said fragment comprises the sequence encoding between one to three of the CDR regions of said human antibody.

69. A vector comprising the nucleic acid molecule, or fragment thereof, according to any one of claims 65-68.

70. The vector according to claim 69, wherein said vector further comprises expression control sequences operably linked to said nucleic acid.

71. A nucleic acid molecule isolated from a non-human animal that encodes a human antibody light chain or the antigen-binding portion thereof that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02).

72. The isolated nucleic acid molecule according to claim 71, wherein said nucleic acid molecule is isolated from a hybridoma that produces said human antibody.

73. An isolated nucleic acid molecule, or a fragment thereof, encoding a human antibody light chain or antigen-binding portion thereof that specifically binds to *Pseudomonas aeruginosa* LPS comprising a nucleotide sequence encoding an amino acid sequence selected from the group consisting of: SEQ ID NO: 22; SEQ ID NO: 23; SEQ ID NO: 24; SEQ ID NO: 25; SEQ ID NO: 26; SEQ ID NO: 27; SEQ ID NO: 28; SEQ ID NO: 29; and SEQ ID NO: 30.

74. The isolated nucleic acid molecule of claim 71 wherein said fragment comprises the sequence encoding between one to three of the CDR regions of said human antibody.

75. A vector comprising the nucleic acid molecule according to any one of claims 71-74.

76. The vector according to claim 75, wherein said vector further comprises an expression control sequence operably linked to said nucleic acid.

77. An isolated host cell comprising

a) a nucleic acid molecule that was isolated from a non-human animal and encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02); or

b) a vector comprising said nucleic acid molecule.

78. An isolated host cell comprising:

a) a nucleic acid molecule that was isolated from a non-human animal and encodes a heavy chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02); or

b) a vector comprising said nucleic acid molecule.

79. An isolated host cell comprising:

a) an nucleic acid molecule that was isolated from a non-human animal and encodes a heavy chain or the antigen-binding portion thereof and an isolated nucleic acid molecule that encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *Pseudomonas*

aeruginosa strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02)
or

b) a vector or vectors comprising said nucleic acid molecules.

80. The isolated host cell of any one of claims 77-79 wherein said cells are chosen from the list consisting of hybridoma cells, bacterial cells, yeast cells, insect cells, amphibian cells and mammalian cells.

81. The host cell according to claim 80, wherein said mammalian cell is selected from the list consisting of a human cell, a mouse cell, a rat cell, a dog cell, a monkey cell, a goat cell, a pig cell, a bovine cell and a hamster cell.

82. The method according to claim 80, wherein said mammalian cells are selected from the list consisting of HeLa cells, NIH 3T3 cells, CHO cells, BHK cells, VERO cells, CV-1 cells, NS/0 cells and COS cells.

83. A method of recombinantly producing the heavy chain or the antigen-binding portion thereof, the light chain or the antigen-binding portion thereof, or both the light chain and heavy chain or antigen-binding portions thereof, of a human antibody that was identified from a non-human animal and specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), comprising the step of cultivating the host cell according to any one of claims 77-81 under conditions in which the nucleic acid molecules are expressed.

84. An isolated heavy chain or antigen-binding portion thereof obtained from the antibody according to claim 1 or claim 2, encoded by the nucleic acid molecule according to claim 67, or isolated from the host cell according to claim 78.

85. An isolated light chain or antigen-binding portion thereof obtained from the antibody according to claim 1 or claim 2, encoded by the nucleic acid molecule according to claim 73, or isolated from the host cell according to claim 77.

86. A non-human transgenic animal comprising the nucleic acid molecule according to claim 67, wherein said non-human transgenic animal expresses said nucleic acid molecule.

87. A non-human transgenic animal comprising the nucleic acid molecule according to claim 73, wherein said non-human transgenic animal expresses said nucleic acid molecule.

88. A non-human transgenic animal comprising an isolated nucleic acid molecule that encodes a heavy chain or the antigen-binding portion thereof and an isolated nucleic acid molecule that encodes a light chain or the antigen-binding portion thereof of a human antibody that specifically binds to LPS from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02), wherein said animal expresses said nucleic acid molecules.

89. The non-human transgenic animal according to any one of claims 86-88, wherein said animal is selected from the list consisting of a mouse, a rat, a hamster, a cow, a sheep, a primate, a horse and a pig.

90. The non-human transgenic animal according to any one of claims 86-88, wherein a human antibody resulting from expression of said isolated nucleic acid molecules or portions thereof is expressed on the surface of cells derived from said animal's B lymphocytic cells or progeny thereof.

91. The non-human transgenic animal according to any one of claims 86-88, wherein a human antibody resulting from expression of said isolated nucleic acid

molecules or a portion thereof is secreted into the lymph, blood, milk, saliva, or ascites of said animal.

92. A fusion protein comprising the isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2 and a second polypeptide sequence.

93. The fusion protein of claim 92, wherein said second polypeptide sequence is chosen from the list consisting of an epitope tag, an affinity tag, a toxic polypeptide, an antibiotic, an enzyme, a second antibody sequence, a complement protein, and a cytokine.

94. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein the heavy chain isotype of said antibody is mu, gamma, delta, epsilon or alpha.

95. The isolated heavy chain or antigen-binding portion thereof according to claim 84 comprising between one to ten amino acid substitutions that increase the serum half-life of said antibody.

96. The isolated light chain or antigen-binding portion thereof according to claim 85 comprising between one to ten amino acid substitutions that increase the serum half-life of said antibody.

97. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein the antibody or portion thereof is produced by a process comprising the steps of:

a) immunizing a non-human animal comprising a human immunoglobulin locus with an antigen selected from the group consisting of an isolated *Pseudomonas aeruginosa* LPS preparation, a virulent *Pseudomonas aeruginosa* cell preparation, an attenuated *Pseudomonas aeruginosa* cell preparation, and a killed *Pseudomonas aeruginosa* cell preparation;

- b) allowing said non-human animal to mount an immune response to said antigen; and
- c) isolating said antibody from said non-human animal.

98. The isolated human antibody or antigen-binding portion thereof according to claim 1 or claim 2, wherein said antibody or portion thereof is isolated from an animal or cell that is free of contaminating human biomaterials.

99. The isolated human antibody or antigen-binding portion thereof according to claim 99 wherein said biomaterials are chosen from the list consisting of viruses, enzymes, hormones, cytokines, receptors, receptor ligands, immunoglobulins, complement, nuclear proteins, and cytoplasmic signaling proteins.

100. The isolated human antibody or antigen-binding portion thereof according to claim 100 wherein said viruses are Epstein-Barr virus or retroviruses.

101. A hybridoma cell line having American Type Culture Collection Deposit Designation PTA-5384, PTA-5385 or PTA-5386.

102. A monoclonal antibody produced by the hybridoma cell line according to claim 101.

103. An isolated human antibody or an antigen-binding portion thereof, that specifically binds LPS O-specific side chain from one of *Pseudomonas aeruginosa* strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02).

104. The antibody antigen-binding portion according to claim 103, which is monoclonal.

105. The antibody or antigen-binding portion according to claim 103 or 104, having a relative binding avidity of about 1.0.

106. The antibody or antigen-binding portion according to claim 104, that specifically binds the LPS O-specific side chain of one strain and does not bind to the LPS O-specific side chain of any other of the listed strains.

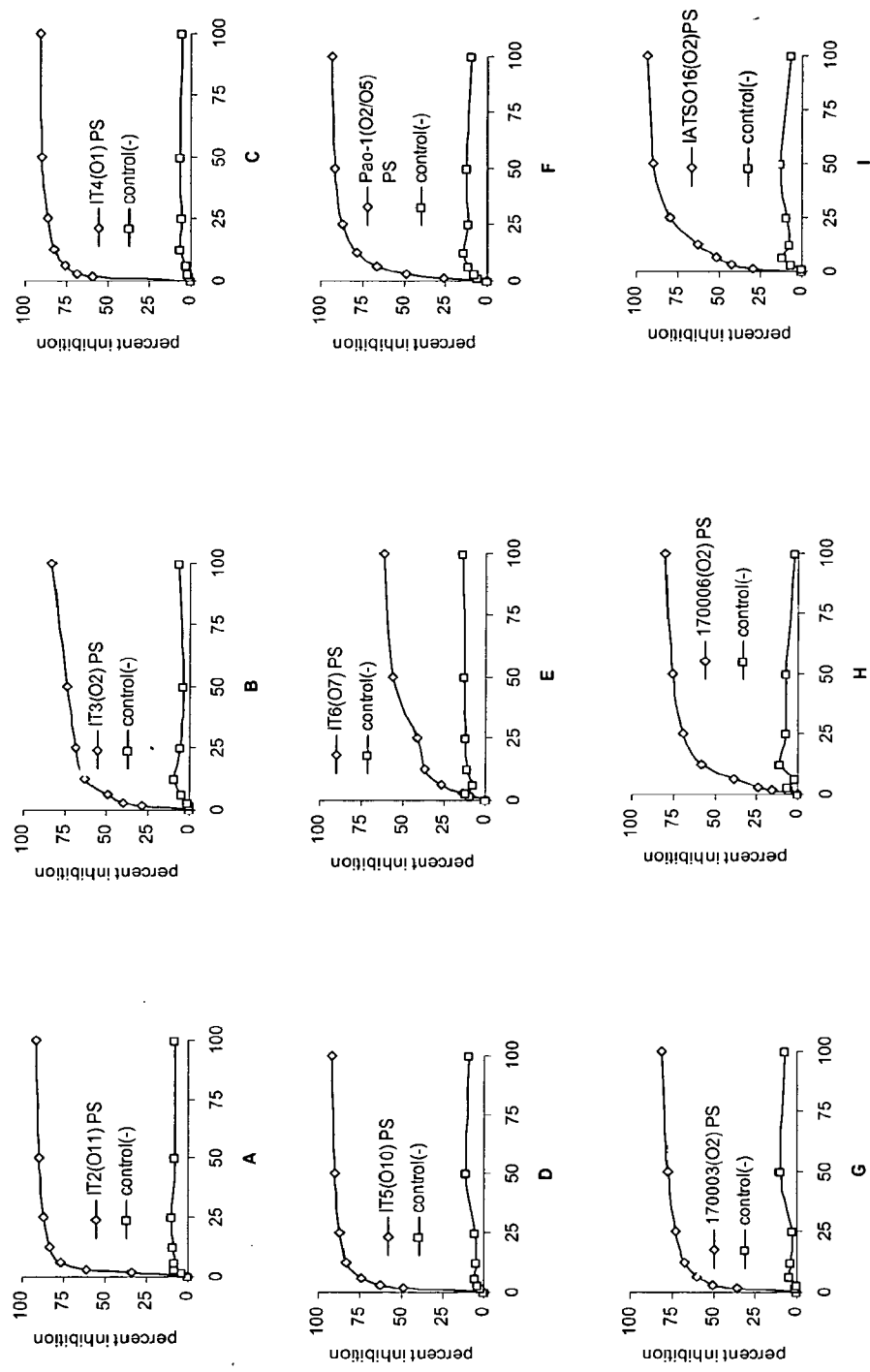
107. A passive vaccine for preventing or inhibiting *Pseudomonas aeruginosa* infection comprising one or more human monoclonal antibodies or an antigen-binding portion thereof, selected from the group consisting of the antibody or portion according to claim 1 or 102.

108. The passive vaccine according to claim 107 comprising two or more human monoclonal antibodies or an antigen-binding portion thereof, wherein said human monoclonal antibodies or portions specifically bind different strains of *Pseudomonas aeruginosa*.

109. A method for detecting the presence of *Pseudomonas aeruginosa* in a biological sample comprising the steps of contacting said sample with an antibody or antigen-binding portion thereof according to any one of claims 1 or 102.

Abstract

The invention described herein provides for human antibodies produced in non-human animals that specifically bind to lipopolysaccharide (LPS) from strains Fisher Devlin (International Serogroups) It-2 (011), It-3 (02), It-4 (01), It-5 (010), It-6 (07), PA01 (05), 170003 (02), IATS016 (02/05), and 170006 (02). The invention further provides methods for making the antibodies in a non-human animal, expression of the antibodies in cell lines including hybridomas and recombinant host cell systems. Also provided are kits and pharmaceutical compositions comprising the antibodies and methods of treating or preventing pseudomonas infection by administering to a patient the pharmaceutical compositions described herein.



μg/ml of PS added to reaction mixture

Figure 1

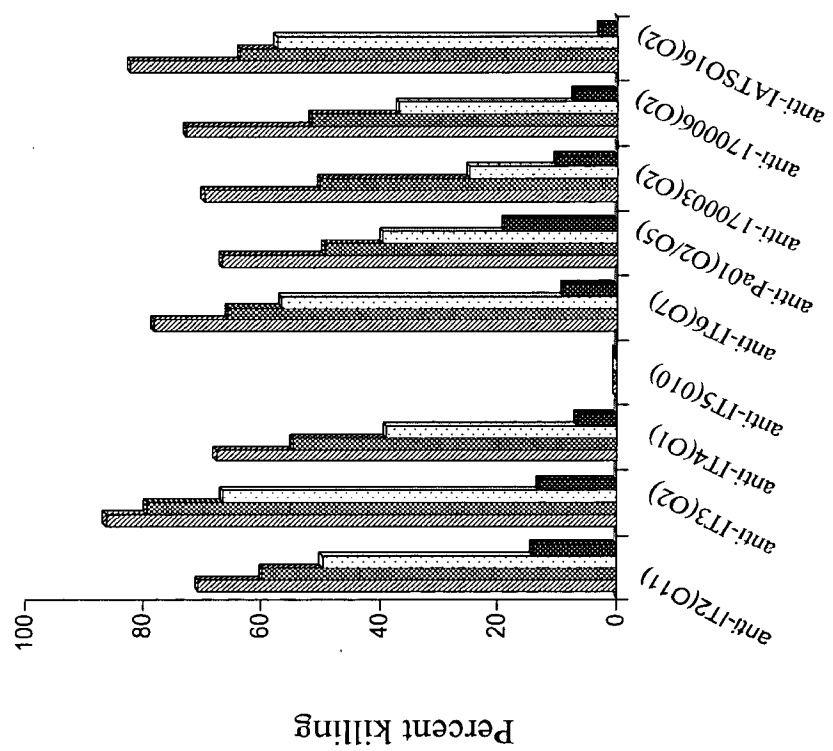


Figure 2

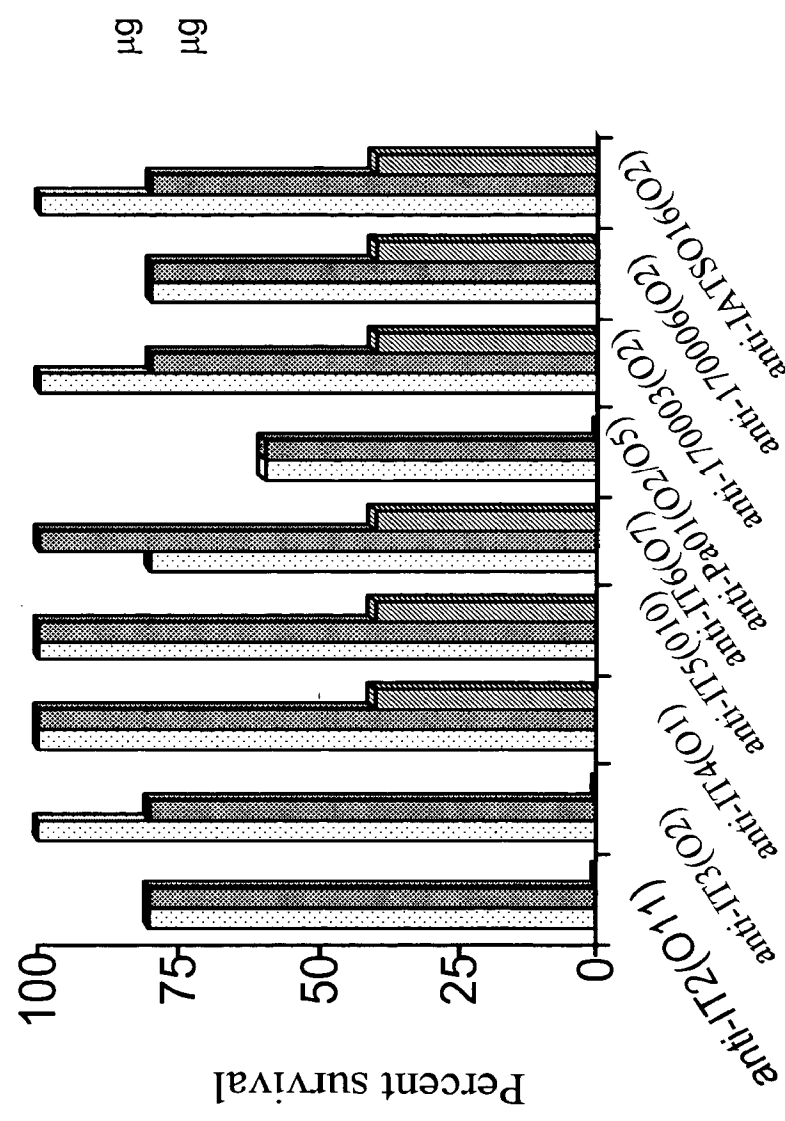


Figure 3

INVENTOR INFORMATION

Inventor One Given Name:: John R
Family Name:: Schreiber
Postal Address Line One:: 544 Battles Road
City:: Gates Mills
State or Province:: OH
Country:: USA
Postal or Zip Code:: 44040
Citizenship Country:: USA

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APPLICATION INFORMATION

Title Line One:: HUMAN ANTI-PSEUDOMONAS-AERUGINOSA ANTIBO
Title Line Two:: DIES DERIVED FROM TRANSGENIC XENOMOUSE
Total Drawing Sheets:: 3
Formal Drawings?:: No
Application Type:: Provisional
Docket Number:: ABX-CW2 PROV
Secrecy Order in Parent Appl.?:: No

REPRESENTATIVE INFORMATION

Representative Customer Number:: 1473
Registration Number One:: 27794
Registration Number Two:: 38479
Registration Number Three:: 48314

Source:: PrintEFS Version 1.0.1